

# Package ‘BaalChIP’

May 1, 2026

**Title** BaalChIP: Bayesian analysis of allele-specific transcription factor binding in cancer genomes

**Description** The package offers functions to process multiple ChIP-seq BAM files and detect allele-specific events. Computes allele counts at individual variants (SNPs/SNVs), implements extensive QC steps to remove problematic variants, and utilizes a bayesian framework to identify statistically significant allele-specific events. BaalChIP is able to account for copy number differences between the two alleles, a known phenotypical feature of cancer samples.

**Version** 1.39.0

**Author** Ines de Santiago, Wei Liu, Ke Yuan, Martin O'Reilly, Chandra SR Chilamakuri, Bruce Ponder, Kerstin Meyer, Florian Markowetz

**Maintainer** Ines de Santiago <inesdesantiago@gmail.com>

**Depends** R (>= 3.3.1), GenomicRanges, IRanges, Rsamtools,

**Imports** GenomicAlignments, GenomeInfoDb, doParallel, parallel, doBy, reshape2, scales, coda, foreach, ggplot2, methods, utils, graphics, stats

**Suggests** RUnit, BiocGenerics, knitr, rmarkdown, BiocStyle

**VignetteBuilder** knitr

**License** Artistic-2.0

**LazyData** true

**RoxygenNote** 7.1.1

**biocViews** Software, ChIPSeq, Bayesian, Sequencing

**git\_url** <https://git.bioconductor.org/packages/BaalChIP>

**git\_branch** devel

**git\_last\_commit** c73e9ab

**git\_last\_commit\_date** 2026-04-28

**Repository** Bioconductor 3.24

**Date/Publication** 2026-05-01

## Contents

adjustmentBaalPlot . . . . .	2
alleleCounts . . . . .	3
BaalChIP . . . . .	4
BaalChIP.get . . . . .	5
BaalChIP.report . . . . .	6
BaalChIP.run . . . . .	8
BaalObject . . . . .	9
blacklist_hg19 . . . . .	9
ENCODEexample . . . . .	10
FAIREexample . . . . .	10
filter1allele . . . . .	11
filterIntbias . . . . .	12
getASB . . . . .	13
mergePerGroup . . . . .	15
pickrell2011cov1_hg19 . . . . .	16
plotQC . . . . .	17
plotSimul . . . . .	18
QCfilter . . . . .	18
summaryASB . . . . .	19
summaryQC . . . . .	20
UniqueMappability50bp_hg19 . . . . .	21
<b>Index</b>	<b>22</b>

---

adjustmentBaalPlot      *Method adjustmentBaalPlot*

---

### Description

Produces a density plot of the distribution of allelic ratios (REF/TOTAL) before and after BaalChIP adjustment for RM and RAF biases.

### Usage

```
adjustmentBaalPlot(.Object, col = c("green3", "gray50"))

## S4 method for signature 'BaalChIP'
adjustmentBaalPlot(.Object, col = c("green3", "gray50"))
```

### Arguments

.Object      An object of the [BaalChIP](#) class.

col          A character vector indicating the colours for the density plot ( default is c('green3','gray50') ).

### Value

A plot

**Author(s)**

Ines de Santiago

**See Also**

[BaalChIP.report, summaryQC](#)

**Examples**

```
data('BaalObject')
adjustmentBaalPlot(BaalObject)
adjustmentBaalPlot(BaalObject, col=c('blue', 'pink'))
```

---

alleleCounts	<i>Method alleleCounts</i>
--------------	----------------------------

---

**Description**

Generates allele-specific read count data from each BAM ChIP-seq dataset for each variant.

**Usage**

```
alleleCounts(.Object, min_base_quality = 10, min_mapq = 15, verbose = TRUE)

## S4 method for signature 'BaalChIP'
alleleCounts(.Object, min_base_quality = 10, min_mapq = 15, verbose = TRUE)
```

**Arguments**

.Object	An object of the <a href="#">BaalChIP</a> class.
min_base_quality	A numeric value indicating the minimum read base quality below which the base is ignored when summarizing pileup information (default 10).
min_mapq	A numeric value indicating the minimum mapping quality (MAPQ) below which the entire read is ignored (default 15).
verbose	logical. If TRUE reports extra information on the process

**Details**

Utilizes the information within the `samples` slot of a `BaalChIP` object. Will primarily find all variants overlapping peaks. Then, for each variant, computes the number of reads carrying the reference (REF) and alternative (ALT) alleles.

**Value**

An updated [BaalChIP](#) object with the slot `alleleCounts` containing a list of `GRanges` objects.

**Note**

`BaalChIP` computes allelic counts at each variant position with `Rsamtools` `pileup` function. The algorithm follows `pileup::Rsamtools` by automatically excluding reads flagged as unmapped, secondary, duplicate, or not passing quality controls.

**Author(s)**

Ines de Santiago

**See Also**

[BaalChIP.get](#)

**Examples**

```

samplesheet <- system.file("test", "exampleChIP.tsv", package = "BaalChIP")
hets <- c("MCF7"= system.file("test", "MCF7_hetSNP.txt", package = "BaalChIP"),
        "GM12891"= system.file("test", "GM12891_hetSNP.txt", package = "BaalChIP"))
res <- BaalChIP(samplesheet=samplesheet, hets=hets)
res <- alleleCounts(res, min_base_quality=10, min_mapq=15)

#retrieve alleleCounts:
counts <- BaalChIP.get(res, 'alleleCountsPerBam')

#alleleCounts are grouped by bam_name and group_name:
names(counts)
names(counts[['MCF7']])

#check out the result for one of the bam files:
counts[['MCF7']][[1]]

```

---

BaalChIP

*BaalChIP-class*

---

**Description**

This S4 class includes a series of methods for detecting allele-specific events from multiple ChIP-seq datasets.

**Usage**

```
BaalChIP(samplesheet = NULL, hets = NULL, CorrectWithgDNA = list())
```

**Arguments**

samplesheet	<p>A character string indicating the filename for a .tsv file. Column names in the .tsv file should include:</p> <ul style="list-style-type: none"> <li>• group_name: identifier string to group samples together</li> <li>• target: identifier string for factor (transcription factor, protein)</li> <li>• replicate_number: replicate number of sample</li> <li>• bam_name: file path for BAM file containing aligned reads for ChIP sample. If duplicated reads are flagged they will not be included in the allelic count data</li> <li>• bed_name: path for BED file containing peaks for ChIP sample</li> <li>• SampleID: identifier string for sample. If not given will use &lt;group_name&gt;_&lt;target&gt;_&lt;replicate&gt;</li> </ul>
hets	<p>A named vector with filenames for the .txt variant files to be used. The names in the vector should correspond to group_name strings in the .tsv samplesheet. Columns names in the .txt file should include:</p>

- ID: unique identifier string per variant. Identifiers have to be unique, and no more than one identifier should be present per data record. If there is no identifier available, then use an arbitrary name to name each variant
- CHROM: chromosome identifier from the reference genome per variant (same genome build as BAM and BED files provided)
- POS: the reference position (1-based)
- REF: reference base. Each base must be one of A,C,G,T in uppercase. Multiple bases are not permitted
- ALT: alternate non-reference base. Each base must be one of A,C,G,T in uppercase. Multiple bases are not permitted
- RAF: [Optional] a value ranging from 0 to 1 for each variant denoting the relative allele frequency (RAF). A value between 0.5 and 1 denotes a bias to the reference allele, and a value between 0 and 0.5 a bias to the alternate allele. If neither RAF or CorrectWithgDNA are given, BaalChIP will not correct for relative allele frequency (copy-number) bias. If both RAF and CorrectWithgDNA are given, BaalChIP will give priority to the RAF values of the 'hets' files and will use these values to correct for relative allele frequency (copy-number) bias.

#### CorrectWithgDNA

An optional named list with complete file paths for the .bam gDNA files to be used. The names in the list should correspond to group\_name strings in the .tsv samplesheet. Allelic read counts from all gDNA files are pooled together to generate the Reference Allelic Ratios (RAF) directly from input data. If missing, BaalChIP will try to read the background allelic ratios from the information in the RAF column of the 'hets' files indicated by the hets parameter. If both RAF and CorrectWithgDNA are missing, BaalChIP will not correct for relative allele frequency (copy-number) bias.

#### Value

.Object An object of the [BaalChIP](#) class.

#### Author(s)

Ines de Santiago, Wei Liu, Ke Yuan, Florian Markowetz

#### Examples

```
samplesheet <- system.file("test", "exampleChIP.tsv", package = "BaalChIP")
hets <- c("MCF7"= system.file("test", "MCF7_hetSNP.txt", package = "BaalChIP"),
        "GM12891"= system.file("test", "GM12891_hetSNP.txt", package = "BaalChIP"))
res <- new("BaalChIP", samplesheet=samplesheet, hets=hets)
res <- BaalChIP(samplesheet=samplesheet, hets=hets)
```

---

BaalChIP.get

*Method BaalChIP.get*

---

#### Description

Get information from individual slots in a BaalChIP object.

**Usage**

```

BaalChIP.get(
  .Object,
  what = c("samples", "param", "alleleCountsPerBam", "mergedCounts", "assayedVar",
           "biasTable")
)

## S4 method for signature 'BaalChIP'
BaalChIP.get(
  .Object,
  what = c("samples", "param", "alleleCountsPerBam", "mergedCounts", "assayedVar")
)

```

**Arguments**

<code>.Object</code>	An object of the <a href="#">BaalChIP</a> class.
<code>what</code>	a single character value specifying which information should be retrieved. Options: 'samples', 'param', 'alleleCountsPerBam', 'mergedCounts', 'assayedVar', 'biasTable'.

**Value**

The slot content from an object of the [BaalChIP](#) class.

**Author(s)**

Ines de Santiago

**Examples**

```

data('BaalObject')

#samples data spreadsheet and hets:
BaalChIP.get(BaalObject,what='samples')

#parameters used within run:
BaalChIP.get(BaalObject,what='param')

#retrieve a GRanges list with allele-specific read counts per BAM file:
counts <- BaalChIP.get(BaalObject,what='alleleCountsPerBam')
counts[['MCF7']][[1]]

#retrieve a data.frame with allele-specific read counts per group:
counts <- BaalChIP.get(BaalObject,what='mergedCounts')
head(counts[[1]])

```

---

BaalChIP.report

*Method BaalChIP.report*

---

**Description**

Generates a data.frame per group with all variants and a label for all identified allele-specific binding (ASB) variants.

## Usage

```
BaalChIP.report(.Object)

## S4 method for signature 'BaalChIP'
BaalChIP.report(.Object)
```

## Arguments

`.Object` An object of the [BaalChIP](#) class.

## Details

The reported data frame contains the following columns:

- ID: unique identifier string per analysed variant.
- CHROM: chromosome identifier from the reference genome per variant.
- POS: the reference position (1-based).
- REF: reference base. Each base must be one of A,C,G,T in uppercase.
- ALT: alternate non-reference base. Each base must be one of A,C,G,T in uppercase.
- REF.counts: pooled counts of all reads with the reference allele.
- ALT.counts: pooled counts of all reads with the non-reference allele.
- Total.counts: pooled counts of all reads (REF + ALT).
- AR: allelic ratio calculated directly from sequencing reads (REF / TOTAL).
- RMBias: numerical value indicating the value estimated and applied by BaalChIP for the reference mapping bias. A value between 0.5 and 1 denotes a bias to the reference allele, and a value between 0 and 0.5 a bias to the alternative allele.
- RAF: numerical value indicating the value applied by BaalChIP for the relative allele frequency (RAF) bias correction. A value between 0.5 and 1 denotes a bias to the reference allele, and a value between 0 and 0.5 a bias to the alternative allele.
- Bayes\_lower: lower interval for the estimated allelic ratio (allelic ratio is given by REF / TOTAL).
- Bayes\_upper: upper interval for the estimated allelic ratio (allelic ratio is given by REF / TOTAL).
- Corrected.AR: average estimated allelic ratio (average between Bayes\_lower and Bayes\_upper). A value between 0.5 and 1 denotes a bias to the reference allele, and a value between 0 and 0.5 a bias to the alternative allele.
- isASB: logical value indicating BaalChIP's classification of variants into allele-specific.

## Value

A named list, with a data.frame per group.

## Author(s)

Ines de Santiago

## See Also

[summaryASB](#), [getASB](#)

## Examples

```
data('BaalObject')
report <- BaalChIP.report(BaalObject)

#the reported list is grouped by group_name:
names(report)

#check out the report for one of the groups:
head(report[['MCF7']])
```

---

BaalChIP.run

*Method BaalChIP.run*

---

## Description

BaalChIP.run is a wrapper convenience function, to compute allele counts and perform quality controls in one step. This function will use the package's defaults.

## Usage

```
BaalChIP.run(.Object, cores = 4, verbose = TRUE)

## S4 method for signature 'BaalChIP'
BaalChIP.run(.Object, cores = 4, verbose = TRUE)
```

## Arguments

.Object	An object of the <a href="#">BaalChIP</a> class.
cores	number of cores for parallel computing (default is 4).
verbose	logical. If TRUE reports extra information on the process

## Details

This function is a wrapper of the following functions: [alleleCounts](#), [QCfilter](#), [mergePerGroup](#), [filter1allele](#), [getASB](#)

## Value

An object of the [BaalChIP](#) class.

## Author(s)

Ines de Santiago

## See Also

[summaryQC](#), [plotQC](#)

**Examples**

```

samplesheet <- system.file("test", "exampleChIP.tsv", package = "BaalChIP")
hets <- c("MCF7"= system.file("test", "MCF7_hetSNP.txt", package = "BaalChIP"),
        "GM12891"= system.file("test", "GM12891_hetSNP.txt", package = "BaalChIP"))
res <- BaalChIP(samplesheet=samplesheet, hets=hets)
res <- BaalChIP.run(res, cores=2)

#summary of the QC step
summaryQC(res)
#summary of the ASB step
summaryASB(res)

```

BaalObject

*BaalObject example dataset***Description**

BaalObject example dataset

**Format**

A BaalChIP-class object

**Author(s)**

Ines de Santiago &lt;ines.desantiago@cruk.cam.ac.uk&gt;

blacklist\_hg19

*Blacklisted genomic regions***Description**

A GRanges object containing blacklisted regions identified by the ENCODE and modENCODE consortia. These correspond to artifact regions that tend to show artificially high signal (excessive unstructured anomalous reads mapping). Selected from mappability track of the UCSC genome browser (hg19, wgEncodeDacMapabilityConsensusExcludable and wgEncodeDukeMapabilityRegionsExcludable tables).

Code used to retrieve these regions:

- `curl http://hgdownload.cse.ucsc.edu/goldenPath/hg19/encodeDCC/wgEncodeMapability/wgEncodeDacMapability > hg19_DACExcludable.txt`
- `cat hg19_DUKEExcludable.txt hg19_DACExcludable.txt | grep -v "^#" | cut -f 2,3,4,5,6,7 | sort -k1,1 -k2,2n | mergeBed -nms -i stdin > hg19_DUKE_DAC.bed`

Used as 'RegionsToFilter' within the QCfilter function so that variants overlapping these regions will be removed.

**Format**

A GRanges object of 1378 ranges.

**Details**

Note that these blacklists are applicable to functional genomic data (e.g. ChIP-seq, MNase-seq, DNase-seq, FAIRE-seq) of short reads (20-100bp reads). These are not applicable to RNA-seq or other transcriptome data types.

**Author(s)**

Ines de Santiago <ines.desantiago@cruk.cam.ac.uk>

---

ENCODEexample      *ENCODEexample example dataset*

---

**Description**

ENCODEexample example dataset

**Format**

A BaalChIP-class object

**Author(s)**

Ines de Santiago <ines.desantiago@cruk.cam.ac.uk>

**Source**

Downloaded from supplementary data of BaalChIP paper

---

FAIREexample      *FAIREexample example dataset*

---

**Description**

FAIREexample example dataset

**Format**

A BaalChIP-class object

**Author(s)**

Ines de Santiago <ines.desantiago@cruk.cam.ac.uk>

**Source**

Downloaded from supplementary data of BaalChIP paper

---

filter1allele	<i>Method filter1allele</i>
---------------	-----------------------------

---

### Description

Filters the data frame available within a [BaalChIP](#) object (slot mergedCounts). This filter ignores variants for which only one allele is observed after pooling ChIP-seq reads from all datasets.

### Usage

```
filter1allele(.Object)

## S4 method for signature 'BaalChIP'
filter1allele(.Object)
```

### Arguments

.Object            An object of the [BaalChIP](#) class.

### Value

An updated [BaalChIP](#) object with the slot mergedCounts containing a data.frame of merged samples per group with variants that pass the filter.

### Author(s)

Ines de Santiago

### See Also

[BaalChIP.get](#), [plotQC](#), [summaryQC](#)

### Examples

```
samplesheet <- system.file("test", "exampleChIP.tsv", package = "BaalChIP")
hets <- c("MCF7"= system.file("test", "MCF7_hetSNP.txt", package = "BaalChIP"),
         "GM12891"= system.file("test", "GM12891_hetSNP.txt", package = "BaalChIP"))
res <- BaalChIP(samplesheet=samplesheet, hets=hets)
res <- alleleCounts(res, min_base_quality=10, min_mapq=15)
data('blacklist_hg19')
data('pickrell12011cov1_hg19')
data('UniqueMappability50bp_hg19')
res <- QCfilter(res,
               RegionsToFilter=list('blacklist'=blacklist_hg19,
                                   'highcoverage'=pickrell12011cov1_hg19),
               RegionsToKeep=list('UniqueMappability'=UniqueMappability50bp_hg19))

res <- mergePerGroup(res)
res <- filter1allele(res)

#retrieve mergedCounts:
counts <- BaalChIP.get(res, 'mergedCounts')
```

```
#mergedCounts are grouped by group_name:
names(counts)
sapply(counts, dim)

#check out the result for one of the groups:
head(counts[[1]])
```

---

filterIntbias	<i>Method filterIntbias</i>
---------------	-----------------------------

---

## Description

Filters the data frame available within a [BaalChIP](#) object (slot `alleleCounts`). This filter performs simulations of reads of the same length as the original ChIP-seq reads, aligns the simulated reads to the genome, calculates the allelic ratios for each variant and finally ignores those variants for which the allelic ratio (REF/TOTAL) is different than 0.5.

## Usage

```
filterIntbias(
  .Object,
  simul_output = NULL,
  tmpfile_prefix = NULL,
  simulation_script = "local",
  alignmentSimulArgs = NULL,
  skipScriptRun = FALSE,
  verbose = TRUE
)

## S4 method for signature 'BaalChIP'
filterIntbias(
  .Object,
  simul_output = NULL,
  tmpfile_prefix = NULL,
  simulation_script = "local",
  alignmentSimulArgs = NULL,
  skipScriptRun = FALSE,
  verbose = TRUE
)
```

## Arguments

<code>.Object</code>	An object of the <a href="#">BaalChIP</a> class.
<code>simul_output</code>	a non-empty character vector giving the directory of where to save the FASTQ and BAM files generated by the function. If <code>NULL</code> , a random directory under the current working directory will be generated.
<code>tmpfile_prefix</code>	an optional character vector giving the initial part of the name of the FASTQ and BAM files generated by the function. If <code>NULL</code> , a random name will be generated.

simulation_script	the file path for simulation script containing the instructions of simulation and alignment commands. If NULL, the default simulation script distributed with BaalChIP ('extra/simulation_run.sh') will be used.
alignmentSimulArgs	a character vector with arguments to the simulation script. If NULL no arguments are passed.
skipScriptRun	a logical value indicating if simulation BAM files should not be generated. If TRUE BaalChIP will look for the BAM files in the 'simul_output/temp_prefix' (default is FALSE).
verbose	logical. If TRUE reports extra information on the process

**Value**

An updated [BaalChIP](#) object with the slot `alleleCounts` containing a list of GRanges objects that pass filters.

**Author(s)**

Ines de Santiago

**See Also**

[BaalChIP.get](#), [plotQC](#), [summaryQC](#)

**Examples**

```

samplesheet <- system.file("test", "exampleChIP.tsv", package = "BaalChIP")
hets <- c("MCF7"= system.file("test", "MCF7_hetSNP.txt", package = "BaalChIP"),
        "GM12891"= system.file("test", "GM12891_hetSNP.txt", package = "BaalChIP"))
res <- BaalChIP(samplesheet=samplesheet, hets=hets)
res <- alleleCounts(res, min_base_quality=10, min_mapq=15)
skipScriptRun=TRUE #For demonstration purposes only (read details in vignette)
res <- filterIntbias(res,
                    simul_output=system.file('test/simuloutput', package='BaalChIP'),
                    tmpfile_prefix='c67c6ec6c433', skipScriptRun=TRUE)

#check results
plotSimul(res)
summaryQC(res)

```

---

getASB

*Method getASB*

---

**Description**

getASB identifies allele-specific binding events using a bayesian framework.

**Usage**

```

getASB(
  .Object,
  Iter = 5000,
  conf_level = 0.95,
  cores = 4,
  RMcorrection = TRUE,
  RAFcorrection = TRUE,
  verbose = TRUE
)

## S4 method for signature 'BaalChIP'
getASB(
  .Object,
  Iter = 5000,
  conf_level = 0.95,
  cores = 4,
  RMcorrection = TRUE,
  RAFcorrection = TRUE,
  verbose = TRUE
)

```

**Arguments**

<code>.Object</code>	An object of the <a href="#">BaalChIP</a> class.
<code>Iter</code>	Maximum number of iterations (default 5000).
<code>conf_level</code>	Confidence interval in the estimated allelic ratio (default 0.95).
<code>cores</code>	number of cores for parallel computing (default is 4).
<code>RMcorrection</code>	Logical value indicating if reference mapping (RM) bias should be applied (default TRUE). If FALSE will not correct for reference allele mapping bias. If TRUE will estimate the RM bias from the overall reference allele proportion.
<code>RAFcorrection</code>	Logical value indicating if relative allele frequency (RAF) bias correction should be applied (default TRUE). If TRUE will read RAF values for each variant from hets files (RAF column name). If FALSE will not correct for relative allele frequency bias.
<code>verbose</code>	logical. If TRUE reports extra information on the process

**Value**

An updated [BaalChIP](#) object with the slot ASB containing variants identified as allele-specific.

**Author(s)**

Wei Liu, Ke Yuan, Ines de Santiago

**See Also**

[summaryASB](#), [BaalChIP.report](#)

## Examples

```
samplesheet <- system.file("test", "exampleChIP.tsv", package = "BaalChIP")
hets <- c("MCF7"= system.file("test", "MCF7_hetSNP.txt", package = "BaalChIP"),
         "GM12891"= system.file("test", "GM12891_hetSNP.txt", package = "BaalChIP"))
res <- BaalChIP(samplesheet=samplesheet, hets=hets)
res <- alleleCounts(res, min_base_quality=10, min_mapq=15)
res <- mergePerGroup(res)
res <- getASB(res, cores=2)

#summary - number of significant ASB variants
summaryASB(res)

#report result
res <- BaalChIP.report(res)
```

---

mergePerGroup	<i>Method mergePerGroup</i>
---------------	-----------------------------

---

## Description

Merges all ChIP-seq datasets within a group of samples creating a data.frame that contains allele-specific read count data for all variants that need to be analysed.

## Usage

```
mergePerGroup(.Object)

## S4 method for signature 'BaalChIP'
mergePerGroup(.Object)
```

## Arguments

.Object            An object of the [BaalChIP](#) class.

## Details

if QCfilter has been applied, will use the most up-to-date variant set available for each individual BAM file (after QC). Missing values are allowed for heterozygous variants that are not available (e.g. do not pass filter for a particular ChIP-seq dataset).

## Value

An updated [BaalChIP](#) object with the slot mergedCounts containing a data.frame of merged samples per group.

## Author(s)

Ines de Santiago

## See Also

[BaalChIP.get](#), [plotQC](#), [summaryQC](#)

**Examples**

```

samplesheet <- system.file("test", "exampleChIP.tsv", package = "BaalChIP")
hets <- c("MCF7"= system.file("test", "MCF7_hetSNP.txt", package = "BaalChIP"),
        "GM12891"= system.file("test", "GM12891_hetSNP.txt", package = "BaalChIP"))
res <- BaalChIP(samplesheet=samplesheet, hets=hets)
res <- alleleCounts(res, min_base_quality=10, min_mapq=15)
data('blacklist_hg19')
data('pickrell2011cov1_hg19')
data('UniqueMappability50bp_hg19')
res <- QCfilter(res,
               RegionsToFilter=list('blacklist'=blacklist_hg19,
                                   'highcoverage'=pickrell2011cov1_hg19),
               RegionsToKeep=list('UniqueMappability'=UniqueMappability50bp_hg19))

res <- mergePerGroup(res)

#retrieve mergedCounts:
counts <- BaalChIP.get(res, 'mergedCounts')

#mergedCounts are grouped by group_name:
names(counts)
sapply(counts, dim)

#check out the result for one of the groups:
head(counts[[1]])

```

---

pickrell2011cov1\_hg19 *Genomic regions of collapsed repeats*

---

**Description**

A GRanges object containing collapsed repeat regions at the 0.1% threshold (hg19 reference). Used as 'RegionsToFilter' within the QCfilter function so that variants overlapping these regions will be removed.

**Format**

A GRanges object of 34359 ranges.

**Author(s)**

Ines de Santiago <ines.desantiago@cruk.cam.ac.uk>

**Source**

File available as supplementary data: Pickrell2011\_seq.cov1.bed (<http://www.ncbi.nlm.nih.gov/pubmed/21690102>)

**References**

Pickrell et al., 2011 (<http://www.ncbi.nlm.nih.gov/pubmed/21690102>)

---

plotQC

*Method plotQC*

---

### Description

Produces different plots of QC results.

### Usage

```
plotQC(.Object, what = "barplot_per_group", addlegend = TRUE, plot = TRUE)

## S4 method for signature 'BaalChIP'
plotQC(.Object, what = "barplot_per_group", addlegend = TRUE, plot = TRUE)
```

### Arguments

.Object	An object of the <a href="#">BaalChIP</a> class.
what	A single character value indicating the type of plot. Options: <ul style="list-style-type: none"><li>• overall_pie: plots the average percentage of variants in each filter category (averaged across all groups analysed).</li><li>• boxplot_per_filter: plots the number of variants that were filtered out per filter category.</li><li>• barplot_per_group: plots the number of variants that were filtered out per group.</li></ul>
addlegend	A logical value indicating if legend should be included in the plot (default TRUE).
plot	a logical value to whether it should plot (TRUE) or not (FALSE). Default is TRUE.

### Value

A plot

### Author(s)

Ines de Santiago

### See Also

[BaalChIP.run](#), [summaryQC](#)

### Examples

```
data('BaalObject')
plotQC(BaalObject, what = 'overall_pie')
plotQC(BaalObject, what = 'boxplot_per_filter', addlegend=FALSE)
plotQC(BaalObject, what = 'barplot_per_group')
```

---

plotSimul                      *Method plotSimul*

---

**Description**

Produces a plot of the proportion of variants that displayed the correct number of mapped simulated reads.

**Usage**

```
plotSimul(.Object, plot = TRUE)

## S4 method for signature 'BaalChIP'
plotSimul(.Object, plot = TRUE)
```

**Arguments**

.Object                      An object of the [BaalChIP](#) class.  
plot                          a logical value to whether it should plot (TRUE) or not (FALSE). Default is TRUE.

**Value**

A plot

**Author(s)**

Ines de Santiago

**Examples**

```
data('BaalObject')
plotSimul(BaalObject)
```

---

QCfilter                      *Method QCfilter*

---

**Description**

Quality control step for removing variants that may be problematic for identification of allele-specific events.

**Usage**

```
QCfilter(.Object, RegionsToFilter = NULL, RegionsToKeep = NULL, verbose = TRUE)

## S4 method for signature 'BaalChIP'
QCfilter(.Object, RegionsToFilter = NULL, RegionsToKeep = NULL, verbose = TRUE)
```

**Arguments**

.Object	An object of the <a href="#">BaalChIP</a> class.
RegionsToFilter	a named list of GRanges objects. Variants overlapping these regions will be removed.
RegionsToKeep	a named list of GRanges objects. Works in an opposite way to 'RegionstoFilter', variants NOT overlapping these regions will be removed.
verbose	logical. If TRUE reports extra information on the process

**Value**

An updated [BaalChIP](#) object with the slot alleleCounts containing a list of GRanges objects that pass filters.

**Author(s)**

Ines de Santiago

**See Also**

[BaalChIP.get](#), [plotQC](#), [summaryQC](#)

**Examples**

```

samplesheet <- system.file("test", "exampleChIP.tsv", package = "BaalChIP")
hets <- c("MCF7"= system.file("test", "MCF7_hetSNP.txt", package = "BaalChIP"),
        "GM12891"= system.file("test", "GM12891_hetSNP.txt", package = "BaalChIP"))
res <- BaalChIP(samplesheet=samplesheet, hets=hets)
res <- alleleCounts(res, min_base_quality=10, min_mapq=15)
data('blacklist_hg19')
data('pickrell2011cov1_hg19')
data('UniqueMappability50bp_hg19')
res <- QCfilter(res,
               RegionsToFilter=list('blacklist'=blacklist_hg19,
                                   'highcoverage'=pickrell2011cov1_hg19),
               RegionsToKeep=list('UniqueMappability'=UniqueMappability50bp_hg19))

#check results
plotQC(res, 'barplot')
summaryQC(res)

```

---

summaryASB

*Method summaryASB*

---

**Description**

Generates summary of ASB test result.

**Usage**

```
summaryASB(.Object)

## S4 method for signature 'BaalChIP'
summaryASB(.Object)
```

**Arguments**

`.Object` An object of the [BaalChIP](#) class.

**Value**

A matrix containing the total number of allele-specific variants (TOTAL) and the number of variants allele-specific for the reference (REF) and alternate alleles (ALT).

**Author(s)**

Ines de Santiago

**See Also**

[getASB](#), [BaalChIP.report](#)

**Examples**

```
data('BaalObject')
summaryASB(BaalObject)
```

---

summaryQC

*Method summaryQC*


---

**Description**

Generates summary of QC result.

**Usage**

```
summaryQC(.Object)

## S4 method for signature 'BaalChIP'
summaryQC(.Object)
```

**Arguments**

`.Object` An object of the [BaalChIP](#) class.

**Value**

A list with two elements:

- `filtering_stats` containing the number of variants that were filtered out in each filter category and the total number that 'pass' all filters.
- `average_stats` containing the average number and average percentage of variants in each filter category, averaged across all analysed groups.

**Author(s)**

Ines de Santiago

**See Also**

[BaalChIP.run](#), [plotQC](#)

**Examples**

```
data('BaalObject')
summaryQC(BaalObject)
```

---

UniqueMappability50bp\_hg19

*Genomic regions of unique mappability*

---

**Description**

A GRanges object containing unique regions with genomic mappability score equal to 1. Selected from DUKE uniqueness mappability track of the UCSC genome browser (hg19, wgEncodeCrgMapabilityAlign50mer table).

Code used to retrieve these regions:

- `curl http://hgdownload.cse.ucsc.edu/gbdb/hg19/bbi/wgEncodeCrgMapabilityAlign50mer.bw > wgEncodeCrgMapabilityAlign50mer.bw`
- `./bigWigToBedGraph wgEncodeCrgMapabilityAlign50mer.bw wgEncodeCrgMapabilityAlign50mer.bedgraph`
- `awk ' if ($4 >= 1) print $0 ' wgEncodeCrgMapabilityAlign50mer.bedgraph > wgEncodeCrgMapabilityAlign50mer_UNIQUEregions.bedgraph`

Used as 'RegionsToKeep' within the QCfilter function so that variants NOT overlapping these regions will be removed.

**Format**

A GRanges object of 9831690 ranges.

**Details**

These regions are not applicable to longer reads (> 50bp).

**Author(s)**

Ines de Santiago <ines.desantiago@cruk.cam.ac.uk>

**Source**

Downloaded from <http://hgdownload.cse.ucsc.edu/gbdb/hg19/bbi/wgEncodeCrgMapabilityAlign50mer.bw>.

# Index

- \* **BaalObject**
  - BaalObject, [9](#)
- \* **ENCODEexample**
  - ENCODEexample, [10](#)
- \* **FAIREexample**
  - FAIREexample, [10](#)
- \* **UniqueMappability50bp\_hg19**
  - UniqueMappability50bp\_hg19, [21](#)
- \* **blacklist\_hg19**
  - blacklist\_hg19, [9](#)
- \* **pickrell2011cov1\_hg19**
  - pickrell2011cov1\_hg19, [16](#)
  
- adjustmentBaalPlot, [2](#)
- adjustmentBaalPlot, BaalChIP-method (adjustmentBaalPlot), [2](#)
- alleleCounts, [3, 8](#)
- alleleCounts, BaalChIP-method (alleleCounts), [3](#)
  
- BaalChIP, [2, 3, 4, 5–8, 11–15, 17–20](#)
- BaalChIP.get, [4, 5, 11, 13, 15, 19](#)
- BaalChIP.get, BaalChIP-method (BaalChIP.get), [5](#)
- BaalChIP.report, [3, 6, 14, 20](#)
- BaalChIP.report, BaalChIP-method (BaalChIP.report), [6](#)
- BaalChIP.run, [8, 17, 21](#)
- BaalChIP.run, BaalChIP-method (BaalChIP.run), [8](#)
- BaalObject, [9](#)
- blacklist\_hg19, [9](#)
  
- ENCODEexample, [10](#)
  
- FAIREexample, [10](#)
- filter1allele, [8, 11](#)
- filter1allele, BaalChIP-method (filter1allele), [11](#)
- filterIntbias, [12](#)
- filterIntbias, BaalChIP-method (filterIntbias), [12](#)
  
- getASB, [7, 8, 13, 20](#)
- getASB, BaalChIP-method (getASB), [13](#)
  
- mergePerGroup, [8, 15](#)
- mergePerGroup, BaalChIP-method (mergePerGroup), [15](#)
  
- pickrell2011cov1\_hg19, [16](#)
- plotQC, [8, 11, 13, 15, 17, 19, 21](#)
- plotQC, BaalChIP-method (plotQC), [17](#)
- plotSimul, [18](#)
- plotSimul, BaalChIP-method (plotSimul), [18](#)
  
- QCfilter, [8, 18](#)
- QCfilter, BaalChIP-method (QCfilter), [18](#)
  
- summaryASB, [7, 14, 19](#)
- summaryASB, BaalChIP-method (summaryASB), [19](#)
- summaryQC, [3, 8, 11, 13, 15, 17, 19, 20](#)
- summaryQC, BaalChIP-method (summaryQC), [20](#)
  
- UniqueMappability50bp\_hg19, [21](#)