

# Package ‘MultipleAlignment’

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**Title** Representation of multiple sequence alignments in Bioconductor

**Description** The package implements a set of S4 classes (DNAMultipleAlignment, RNAMultipleAlignment, AAMultipleAlignment) for representing Multiple Sequence Alignments (MSA). The classes allow users to represent groups of aligned DNA, RNA or amino acid sequences as a single object.

The package also provides functions to read/write such object from/to traditional MSA file formats including Stockholm and Clustal.

**biocViews** Alignment, MultipleSequenceAlignment, Genetics, DataImport, DataRepresentation, Infrastructure

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**BugReports** <https://github.com/Bioconductor/MultipleAlignment/issues>

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detail	<i>Show (display) detailed object content</i>
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### Description

This is a variant of [show](#), offering a more detailed display of object content.

### Usage

```
detail(x, ...)
```

### Arguments

x	An object. The default simply invokes <a href="#">show</a> .
...	Additional arguments. The default definition makes no use of these arguments.

### Value

None; the function is invoked for its side effect (detailed display of object content).

### Author(s)

Martin Morgan

### Examples

```
origMAlign <-
  readDNAMultipleAlignment(filepath =
    system.file("extdata",
                "msx2_mRNA.aln",
                package="MultipleAlignment"),
    format="clustal")
detail(origMAlign)
```

---

MultipleAlignment-class

*MultipleAlignment objects*


---

### Description

The MultipleAlignment class is a container for storing multiple sequence alignments.

### Usage

```
## Constructors:
DNAMultipleAlignment(x=character(), start=NA, end=NA, width=NA,
  use.names=TRUE, rowmask=NULL, colmask=NULL)
RNAMultipleAlignment(x=character(), start=NA, end=NA, width=NA,
  use.names=TRUE, rowmask=NULL, colmask=NULL)
AAMultipleAlignment(x=character(), start=NA, end=NA, width=NA,
  use.names=TRUE, rowmask=NULL, colmask=NULL)

## ... and more (see below)
```

### Arguments

x	Either a character vector (with no NAs), or an <a href="#">XString</a> , <a href="#">XStringSet</a> or <a href="#">XStringViews</a> object containing strings with the same number of characters.
start, end, width	Either NA, a single integer, or an integer vector of the same length as x specifying how x should be "narrowed" (see <a href="#">?narrow</a> in the <b>IRanges</b> package for the details).
use.names	TRUE or FALSE. Should names be preserved?
rowmask	a <a href="#">NormalIRanges</a> object that will set masking for rows
colmask	a <a href="#">NormalIRanges</a> object that will set masking for columns

### Details

The MultipleAlignment class is designed to hold and represent multiple sequence alignments. The rows and columns within an alignment can be masked for ad hoc analyses.

### Value

Each constructor function returns a MultipleAlignment derivative of the same class as the name of the function.

### Accessor methods

In the code snippets below, x is a MultipleAlignment object.

`unmasked(x)`: The underlying [XStringSet](#) object containing the multiple sequence alignment.

`rownames(x)`: NULL or a character vector of the same length as x containing a short user-provided description or comment for each sequence in x.

`rowmask(x)`, `rowmask(x, append, invert) <- value`: Gets and sets the [NormalIRanges](#) object representing the masked rows in `x`. The `append` argument takes `union`, `replace` or `intersect` to indicate how to combine the new value with `rowmask(x)`. The `invert` argument takes a logical argument to indicate whether or not to invert the new mask. The value argument can be of any class that is coercible to a [NormalIRanges](#) via the `as` function.

`colmask(x)`, `colmask(x, append, invert) <- value`: Gets and sets the [NormalIRanges](#) object representing the masked columns in `x`. The `append` argument takes `union`, `replace` or `intersect` to indicate how to combine the new value with `colmask(x)`. The `invert` argument takes a logical argument to indicate whether or not to invert the new mask. The value argument can be of any class that is coercible to a [NormalIRanges](#) via the `as` function.

`maskMotif(x, motif, min.block.width=1, ...)`: Returns a [MultipleAlignment](#) object with a modified column mask based upon motifs found in the consensus string where the consensus string keeps all the columns but drops the masked rows.

**motif** The motif to mask.

**min.block.width** The minimum width of the blocks to mask.

... Additional arguments for `matchPattern`.

`maskGaps(x, min.fraction, min.block.width)`: Returns a [MultipleAlignment](#) object with a modified column mask based upon gaps in the columns. In particular, this mask is defined by `min.block.width` or more consecutive columns that have `min.fraction` or more of their non-masked rows containing gap codes.

**min.fraction** A value in  $[0, 1]$  that indicates the minimum fraction needed to call a gap in the consensus string (default is 0.5).

**min.block.width** A positive integer that indicates the minimum number of consecutive gaps to mask, as defined by `min.fraction` (default is 4).

`nrow(x)`: Returns the number of sequences aligned in `x`.

`ncol(x)`: Returns the number of characters for each alignment in `x`.

`dim(x)`: Equivalent to `c(nrow(x), ncol(x))`.

`maskednrow(x)`: Returns the number of masked aligned sequences in `x`.

`maskedncol(x)`: Returns the number of masked aligned characters in `x`.

`maskeddim(x)`: Equivalent to `c(maskednrow(x), maskedncol(x))`.

`maskedratio(x)`: Equivalent to `maskeddim(x) / dim(x)`.

`nchar(x)`: Returns the number of unmasked aligned characters in `x`, i.e. `ncol(x) - maskedncol(x)`.

`alphabet(x)`: Equivalent to `alphabet(unmasked(x))`.

## Coercion

In the code snippets below, `x` is a [MultipleAlignment](#) object.

`as(from, "DNAStringSet")`, `as(from, "RNAStringSet")`, `as(from, "AAStringSet")`, `as(from, "BStringSet")`:  
Creates an instance of the specified [XStringSet](#) object subtype that contains the unmasked regions of the multiple sequence alignment in `x`.

`as.character(x, use.names)`: Convert `x` to a character vector containing the unmasked regions of the multiple sequence alignment. `use.names` controls whether or not `rownames(x)` should be used to set the names of the returned vector (default is TRUE).

`as.matrix(x, use.names)`: Returns a character matrix containing the "exploded" representation of the unmasked regions of the multiple sequence alignment. `use.names` controls whether or not `rownames(x)` should be used to set the row names of the returned matrix (default is TRUE).

**Display**

**The show() method:** The letters in a MultipleAlignment object are colored when displayed by the show() method. Set global option Biostrings.coloring to FALSE to turn off this coloring.

**The detail() method:** In addition to a show() method, a detail() method is provided (detail() is a new generic function defined in this package, see [?detail](#)).

detail(x, invertColMask, hideMaskedCols): Allows for a full pager driven display of the object so that masked cols and rows can be removed and the entire sequence can be visually inspected. If hideMaskedCols is set to its default value of TRUE then the output will hide all the masked columns in the output. Otherwise, all columns will be displayed along with a row to indicate the masking status. If invertColMask is TRUE then any displayed mask will be flipped so as to represent things in a way consistent with Phylip style files instead of the mask that is actually stored in the MultipleAlignment object. Please notice that invertColMask will be ignored if hideMaskedCols is set to its default value of TRUE since in that case it will not make sense to show any masking information in the output. Masked rows are always hidden in the output.

**Author(s)**

P. Aboyoun and M. Carlson

**See Also**

[MultipleAlignment-IO](#), [MultipleAlignment-utils](#), [XStringSet-class](#), [MaskedXString-class](#)

**Examples**

```
## create an object from file
origMAlign <-
  readDNAMultipleAlignment(filepath =
                           system.file("extdata",
                                         "msx2_mRNA.aln",
                                         package="MultipleAlignment"),
                           format="clustal")

## list the names of the sequences in the alignment
rownames(origMAlign)

## rename the sequences to be the underlying species for MSX2
rownames(origMAlign) <- c("Human", "Chimp", "Cow", "Mouse", "Rat",
                          "Dog", "Chicken", "Salmon")
origMAlign

## See a detailed pager view
if (interactive()) {
  detail(origMAlign)
}

## operations to mask rows
## For columns, just use colmask() and do the same kinds of operations
rowMasked <- origMAlign
rowmask(rowMasked) <- IRanges(start=1, end=3)
rowMasked

## remove rowumn masks
rowmask(rowMasked) <- NULL
```

```

rowMasked

## "select" rows of interest
rowmask(rowMasked, invert=TRUE) <- IRanges(start=4,end=7)
rowMasked

## or mask the rows that intersect with masked rows
rowmask(rowMasked, append="intersect") <- IRanges(start=1,end=5)
rowMasked

## TATA-masked
tataMasked <- maskMotif(origMAlign, "TATA")
colmask(tataMasked)

## automatically mask rows based on consecutive gaps
autoMasked <- maskGaps(origMAlign, min.fraction=0.5, min.block.width=4)
colmask(autoMasked)
autoMasked

## cluster the masked alignments
library(pwalign) # for stringDist()
sdist <- stringDist(as(autoMasked, "DNAStrngSet"), method="hamming")
clust <- hclust(sdist, method = "single")
plot(clust)
fourgroups <- cutree(clust, 4)
fourgroups

```

---

MultipleAlignment-IO *Read/write MultipleAlignment objects*

---

## Description

Functions to read/write [MultipleAlignment](#) objects.

## Usage

```

## Read functions:
readDNAMultipleAlignment(filepath, format)
readRNAMultipleAlignment(filepath, format)
readAAMultipleAlignment(filepath, format)

## Write funtions:
write.phylip(x, filepath)

```

## Arguments

x	A <a href="#">MultipleAlignment</a> object
filepath	A character vector (of arbitrary length when reading, of length 1 when writing) containing the paths to the files to read or write. Note that special values like "" or " cmd" (typically supported by other I/O functions in R) are not supported here. Also filepath cannot be a connection.
format	Either "fasta" (the default), "stockholm", "phylip", or "clustal".

**Value**

Each read function returns a MultipleAlignment derivative of the class that matches the name of the function.

**Author(s)**

P. Aboyoun and M. Carlson

**See Also**

[MultipleAlignment](#)

**Examples**

```
example(MultipleAlignment) # make MultipleAlignment object 'autoMasked'
autoMasked

## write out the alignment object (with current masks) to Phylip format
write.phylip(autoMasked, filepath=tempfile())
```

---

MultipleAlignment-utils

*MultipleAlignment utilities*

---

**Description**

A small set of convenient utilities function to operate on a [MultipleAlignment](#) object.

**Details**

In the code snippets below, x is a [MultipleAlignment](#) object.

`consensusMatrix(x, as.prob, baseOnly)`: Creates an integer matrix containing the column frequencies of the underlying alphabet with masked columns being represented with NA values. If `as.prob` is TRUE, then probabilities are reported, otherwise counts are reported (the default). If `baseOnly` is TRUE, then the non-base letters are collapsed into an "other" category.

`consensusString(x, ...)`: Creates a consensus string for x with the symbol "#" representing a masked column. See [consensusString](#) in the **Biostrings** package for details on the arguments.

`consensusViews(x, ...)`: Similar to the `consensusString` method. It returns a [XStringViews](#) on the consensus string containing subsequence contigs of non-masked columns. Unlike the `consensusString` method, the masked columns in the underlying string contain a consensus value rather than the "#" symbol.

`alphabetFrequency(x, as.prob, collapse)`: Creates an integer matrix containing the row frequencies of the underlying alphabet. If `as.prob` is TRUE, then probabilities are reported, otherwise counts are reported (the default). If `collapse` is TRUE, then returns the overall frequency instead of the frequency by row.

**Value**

See description of each method above for what they return.

**Author(s)**

P. Aboyoun and M. Carlson

**See Also**

[MultipleAlignment](#)

**Examples**

```
example(MultipleAlignment) # make MultipleAlignment object 'autoMasked'  
autoMasked  
  
## calculate frequencies  
alphabetFrequency(autoMasked)  
consensusMatrix(autoMasked, baseOnly=TRUE)[, 84:90]  
  
## get consensus values  
consensusString(autoMasked)  
consensusViews(autoMasked)
```

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