

# Package ‘monaLisa’

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**Type** Package

**Title** Binned Motif Enrichment Analysis and Visualization

**Version** 1.15.0

**Description** Useful functions to work with sequence motifs in the analysis of genomics data. These include methods to annotate genomic regions or sequences with predicted motif hits and to identify motifs that drive observed changes in accessibility or expression. Functions to produce informative visualizations of the obtained results are also provided.

**Depends** R (>= 4.1)

**Imports** BiocGenerics, BiocParallel, Biostrings, BSgenome, circlize, ComplexHeatmap (>= 2.11.1), GenomeInfoDb, GenomicRanges, cli, ggplot2, glmnet, grDevices, grid, IRanges, methods, rlang, RSQLite, stabs, stats, SummarizedExperiment, S4Vectors, TFBSTools, tidyr, tools, utils, XVector

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monaLisa-package

*monaLisa - MOTif aNalysis with Lisa.***Description**

**monaLisa** is a collection of tools that simplify motif enrichment analyses in genomic regions of interest.

**Details**

She makes use of her father Homer (<http://homer.ucsd.edu/homer/index.html>) and other algorithms to search for motif hits and look for enriched motifs in sets of genomic regions, compared to all other regions.

Known motifs can for example be obtained from a collection of transcription factor binding site specificities, such as **JASPAR2020**.

**Author(s)**

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**See Also**

Useful links:

- <https://github.com/fmicompbio/monaLisa>
- <https://bioconductor.org/packages/monaLisa/>
- <https://fmicompbio.github.io/monaLisa/>
- Report bugs at <https://github.com/fmicompbio/monaLisa/issues>

`.calcKmerEnrichment`*Calculate k-mer enrichment***Description**

Given sequences, foreground/background labels and weights, calculate the enrichment of each k-mer in foreground compared to background. This function is called by `calcBinnedKmerEnr()` for each bin if `background != "model"`.

The default type of test is "fisher". Alternatively, a binomial test can be used by `test = "binomial"`. Using Fisher's exact test has the advantage that special cases such as zero background counts are handled without ad-hoc adjustments to the k-mer frequencies.

For `test = "fisher"`, `fisher.test` is used with `alternative = "greater"`, making it a one-sided test for enrichment, as is the case with the binomial test.

**Usage**

```
.calcKmerEnrichment(k, df, test = c("fisher", "binomial"), verbose = FALSE)
```

**Arguments**

<b>k</b>	Numeric scalar giving the length of k-mers to analyze.
<b>df</b>	A <code>DataFrame</code> with sequence information as returned by <code>.iterativeNormForKmers()</code> .
<b>test</b>	Type of motif enrichment test to perform.
<b>verbose</b>	A logical scalar. If <code>TRUE</code> , report on progress.

**Details**

The function works in ZOOPS mode, which means only one or zero occurrences of a k-mer are considered per sequence. This is helpful to reduce the impact of simple sequence repeats occurring in few sequences.

**Value**

A `data.frame` containing the motifs as rows and the columns:

**motifName** : the motif name

**logP** : the log p-value for enrichment (natural logarithm). If `test="binomial"` (default), this log p-value is identical to the one returned by Homer.

**sumForegroundWgtWithHits** : the weighted number of k-mer hits in foreground sequences.

**sumBackgroundWgtWithHits** : the weighted number of k-mer hits in background sequences.

**totalWgtForeground** : the total sum of weights of foreground sequences.

**totalWgtBackground** : the total sum of weights of background sequences.

---

`.calcMotifEnrichment`    *Calculate motif enrichment*

---

**Description**

Given motif counts, foreground/background labels and weights for a set of sequences, calculate the enrichment of each motif in foreground compared to background. This function is called by `calcBinnedMotifEnrR()` for each bin.

The default type of test is "fisher", which is also what Homer uses if "-h" is specified for a hypergeometric test. Alternatively, a binomial test can be used by `test = "binomial"` (what Homer does by default). Using Fisher's exact test has the advantage that special cases such as zero background counts are handled without ad-hoc adjustments to the frequencies.

For `test = "fisher"`, `fisher.test` is used with `alternative = "greater"`, making it a one-sided test for enrichment, as is the case with the binomial test.

**Usage**

```
.calcMotifEnrichment(
  motifHitMatrix,
  df,
  test = c("fisher", "binomial"),
  verbose = FALSE
)
```

**Arguments**

<code>motifHitMatrix</code>	Matrix with 0 and 1 entries for absence or presence of motif hits in each sequence.
<code>df</code>	A <code>DataFrame</code> with sequence information as returned by <code>.iterativeNormForKmers()</code> .
<code>test</code>	Type of motif enrichment test to perform.
<code>verbose</code>	A logical scalar. If <code>TRUE</code> , report on progress.

**Value**

A `data.frame` containing the motifs as rows and the columns:

**`motifName`** : the motif name

**`logP`** : the log p-value for enrichment (natural logarithm). If `test="binomial"` (default), this log p-value is identical to the one returned by Homer.

**`sumForegroundWgtWithHits`** : the sum of the weights of the foreground sequences that have at least one instance of a specific motif (ZOOPS mode).

**`sumBackgroundWgtWithHits`** : the sum of the weights of the background sequences that have at least one instance of a specific motif (ZOOPS mode).

**`totalWgtForeground`** : the total sum of weights of foreground sequences.

**`totalWgtBackground`** : the total sum of weights of background sequences.

---

<code>.calculateGCweight</code>	<i>Get background sequence weights for GC bins</i>
---------------------------------	--

---

**Description**

The logic is based on Homer (version 4.11). All sequences binned depending on GC content (GCbreaks). The numbers of foreground and background sequences in each bin are counted, and weights for background sequences in bin *i* are defined as:  $\text{weight}_i = (\text{number\_fg\_seqs}_i / \text{number\_bg\_seqs}_i) * (\text{number\_bg\_seqs\_total} / \text{number\_fg\_seqs\_total})$

**Usage**

```
.calculateGCweight(
  df,
  GCbreaks = c(0.2, 0.25, 0.3, 0.35, 0.4, 0.45, 0.5, 0.6, 0.7, 0.8),
  verbose = FALSE
)
```

**Arguments**

<code>df</code>	A <code>DataFrame</code> with sequence information.
<code>GCbreaks</code>	The breaks between GC bins. The default value is based on the hard-coded bins used in Homer.
<code>verbose</code>	A logical scalar. If <code>TRUE</code> , report on GC weight calculation.

**Value**

A `DataFrame` of the same dimensions as the input `df`, with the columns `GCfrac`, `GCbin` and `GCwgt` filled in with the sequence GC content, assigned GC bins and weights to correct differences in GC distributions between foreground and background sequences.

---

<code>.checkDfValidity</code>	<i>Check if seqinfo DataFrame is valid</i>
-------------------------------	--

---

### Description

Check if the DataFrame with sequence information is valid, i.e. is of the correct object type (DataFrame) and has all expected columns and attributes.

### Usage

```
.checkDfValidity(df)
```

### Arguments

<code>df</code>	Input object to be checked. It should have an attribute <code>err</code> and columns: <code>seqs</code> : a DNASTringSet object. <code>isForeground</code> that indicates if a sequence is in the foreground group. <code>GCfrac</code> : the fraction of G+C bases per sequence. <code>GCbin</code> : the GC bin for each sequence. <code>GCwgt</code> : the sequence weight to adjust for GC differences between foreground and background sequences. <code>seqWgt</code> : the sequence weight to adjust for k-mer differences between foreground and background sequences.
-----------------	--

### Value

TRUE (invisibly) if `df` is valid, otherwise it raises an exception using `cli::cli_abort()`

---

<code>.checkIfSeqsAreEqualLength</code>	<i>Check if elements of 'x' are have equal lengths</i>
---	--

---

### Description

Check if the elements of 'x' are all equally long. If not, generate a warning.

### Usage

```
.checkIfSeqsAreEqualLength(x)
```

### Arguments

<code>x</code>	An object that implements a width method, typically a GRanges or DNASTringSet object.
----------------	---

### Value

NULL (invisibly). The function is called for its side-effect of generating a warning if elements of the input are not of equal lengths.

---

<code>.cons2matrix</code>	<i>Create matrix from consensus sequence</i>
---------------------------	--

---

### Description

Given a nucleotide sequence of A,C,G,T letter corresponding to a motif's consensus string, construct a positional frequency matrix. This matrix can for example be used as the `profileMatrix` argument in the constructor for a `TFBSTools::PFMatrix` object.

### Usage

```
.cons2matrix(x, n = 100L)
```

### Arguments

<code>x</code>	Character scalar with the motif the consensus sequence.
<code>n</code>	Integer scalar giving the columns sums in the constructed matrix (number of observed bases at each position).

### Value

A positional frequency matrix.

---

<code>.defineBackground</code>	<i>Define background sequence set for a single motif enrichment calculation</i>
--------------------------------	---

---

### Description

Define the background set for the motif enrichment calculation in a single bin, depending on the background mode and given foreground sequences.

### Usage

```
.defineBackground(  
  sqs,  
  bns,  
  bg,  
  currbn,  
  gnm,  
  gnm.regions,  
  gnm.oversample,  
  maxFracN = 0.7,  
  GCbreaks = c(0.2, 0.25, 0.3, 0.35, 0.4, 0.45, 0.5, 0.6, 0.7, 0.8)  
)
```

**Arguments**

<code>sqs, bns, bg</code>	The <code>sqs</code> , <code>bns</code> and background arguments from <code>calcBinnedMotifEnrR</code> .
<code>currbn</code>	An integer scalar with the current bin defining the foreground sequences.
<code>gnm, gnm.regions, gnm.oversample</code>	The <code>genome</code> , <code>genome.regions</code> and <code>genome.oversample</code> arguments from <code>calcBinnedMotifEnrR</code> .
<code>maxFracN</code>	The <code>maxFracN</code> argument from <code>calcBinnedMotifEnrR</code> .
<code>GCbreaks</code>	The breaks between GC bins. The default value is based on the hard-coded bins used in Homer.

**Value**

A `DataFrame` with sequences represented by rows and columns `sqs`, `isForeground`, `GCfrac`, `GCbin`, `GCwgt` and `seqWgt`. Only the first three are already filled in.

---

<code>.filterSeqs</code>	<i>Filter Sequences</i>
--------------------------	-------------------------

---

**Description**

Filter sequences that are unlikely to be useful for motif enrichment analysis. The current defaults are based on HOMER (version 4.11).

**Usage**

```
.filterSeqs(
  seqs,
  maxFracN = 0.7,
  minLength = 5L,
  maxLength = 100000L,
  verbose = FALSE
)
```

**Arguments**

<code>sqs</code>	A <code>DNASTringSet</code> object.
<code>maxFracN</code>	A numeric scalar with the maximal fraction of N bases allowed in a sequence (defaults to 0.7).
<code>minLength</code>	The minimum sequence length (default from Homer). Sequences shorter than this will be filtered out.
<code>maxLength</code>	The maximum sequence length (default from Homer). Sequences bigger than this will be filtered out.
<code>verbose</code>	A logical scalar. If <code>TRUE</code> , report on filtering.

**Details**

The filtering logic is based on `removePoorSeq.pl` from Homer.

**Value**

A logical vector of the same length as `sqs` with `TRUE` indicated to keep the sequence and `FALSE` to filter it out.



---

*.glmnetRandomizedLasso**Randomized Lasso*

---

## Description

This function performs randomized lasso using the `glmnet` package. The function present in the `stabs` package that runs the lasso version was adapted for the randomized lasso here. Randomized lasso stability selection uses this function repeatedly to select predictors.

## Usage

```
.glmnetRandomizedLasso(
  x,
  y,
  q,
  weakness = 1,
  type = c("conservative", "anticonservative"),
  ...
)
```

## Arguments

<code>x</code>	The predictor matrix. Passed to <code>x</code> of <code>glmnet.lasso</code> from <code>stabs</code> package.
<code>y</code>	The response vector. Passed to <code>y</code> of <code>glmnet.lasso</code> from <code>stabs</code> package.
<code>q</code>	The number of variables that are selected on each subsample. Passed to <code>q</code> of <code>glmnet.lasso</code> from <code>stabs</code> package.
<code>weakness</code>	Weakness parameter used in randomized lasso (see details).
<code>type</code>	Parameter passed to <code>type</code> of <code>glmnet.lasso</code> from <code>stabs</code> package. It is a character vector specifying how much the PFER should be controlled. If <code>type</code> is "conservative" (default), then the number of selected variables per subsample is $\leq q$ . If <code>type</code> is "anticonservative" then the number of selected variables per subsample is $\geq q$ .
<code>...</code>	Additional parameters for <code>glmnet</code> .

## Details

This function is identical to `glmnet.lasso` from the `stabs` package. The only addition/modification is the `weakness` parameter which has been added when calling the `glmnet` function by setting `penalty.factor = 1/runif(ncol(x), weakness, 1)`, where `ncol(x)` is the number of predictors.

## Value

The regression output which consists of a list of length 2. The list contains the following:

**selected** - a logical vector of length equal to the total number of predictors. The predictors that were chosen have a value of `TRUE`.

**path** - a logical matrix containing the regularization steps as columns and the predictors as rows. An entry of `TRUE` indicates selection.

**See Also**

[glmnet.lasso](#) and [glmnet](#)

---

```
.iterativeNormForKmers
```

*Adjust for k-mer composition (multiple iterations)*

---

**Description**

Here we run ‘.normForKmers’ multiple times to converge to the final weights that will be used to correct the background sequences for k-mer composition differences compared to the foreground. We closely follow HOMER’s `normalizeSequence()` function found in `Motif2.cpp`. Note that HOMER runs the `normalizeSequence()` one last time after going through all iterations or reaching a low error, which we do not do here.

**Usage**

```
.iterativeNormForKmers(
  df,
  maxKmerSize = 3L,
  minSeqWgt = 0.001,
  maxIter = 160L,
  verbose = FALSE
)
```

**Arguments**

<code>df</code>	A <code>DataFrame</code> with sequence information as returned by <code>.calculateGCweight</code> .
<code>maxKmerSize</code>	Integer scalar giving the maximum k-mer size to consider. The default is set to 3 (like in HOMER), meaning that k-mers of size 1, 2 and 3 are considered.
<code>minSeqWgt</code>	Numeric scalar greater than zero giving the minimal weight of a sequence. The default value (0.001) was also used by HOMER ( <code>HOMER_MINIMUM_SEQ_WEIGHT</code> constant in <code>Motif2.h</code> ).
<code>maxIter</code>	An integer scalar giving the maximum number of times to run <code>.normForKmers</code> . the default is set to 160 (as in HOMER).
<code>verbose</code>	A logical scalar. If <code>TRUE</code> , report on k-mer composition adjustment.

**Value**

A `DataFrame` containing:

**sequenceWeights** : a dataframe containing the sequence GC content, GC bins they were assigned to, the weight to correct for GC differences between foreground and background sequences, the weight to adjust for kmer composition, and the error term

**sequenceNucleotides** : a `DNAStrngSet` object containing the raw sequences

---

.normForKmers	<i>Adjust for k-mer composition (single iteration)</i>
---------------	--

---

## Description

Adjust background sequence weights for differences in k-mer composition compared to the foreground sequences. This function implements a single iteration, and is called iteratively by `.iterativeNormForKmers` to get to the final set of adjusted weights, which will be the result of adjusting for GC and k-mer composition. The logic is based on Homer's `normalizeSequenceIteration()` function found in `Motif2.cpp`.

## Usage

```
.normForKmers(
  kmerFreq,
  goodKmers,
  kmerRC,
  seqWgt,
  isForeground,
  minSeqWgt = 0.001,
  maxSeqWgt = 1000
)
```

## Arguments

kmerFreq	A list with of matrices. The matrix at index <i>i</i> in the list contains the probability of k-mers of length <i>i</i> , for each k-mer (columns) and sequence (rows).
goodKmers	A list of numeric vectors; the element at index <i>i</i> contains the number of good (non-N-containing) k-mers of length <i>i</i> for each sequence.
kmerRC	A list of character vectors; the element at index <i>i</i> contains the reverse complement sequences of all k-mers of length <i>i</i> .
seqWgt	A numeric vector with starting sequence weights at the beginning of the iteration.
isForeground	Logical vector of the same length as <code>seqs</code> . TRUE indicates that the sequence is from the foreground, FALSE that it is a background sequence.
minSeqWgt	Numeric scalar greater than zero giving the minimal weight of a sequence. The default value (0.001) is based on Homer ( <code>HOMER_MINIMUM_SEQ_WEIGHT</code> constant in <code>Motif2.h</code> ).
maxSeqWgt	Numeric scalar greater than zero giving the maximal weight of a sequence. The default value (1000) is based on HOMER ( <code>1 / HOMER_MINIMUM_SEQ_WEIGHT</code> constant in <code>Motif2.h</code> ).

## Value

A named list with elements `seqWgt` (updated weights) and `err` (error measuring difference of foreground and weighted background sequence compositions).

---

annoSeqlogo

---

*Sequence logo annotation*

---

**Description**

Create an annotation for a [Heatmap](#) containing sequence logos.

**Usage**

```
annoSeqlogo(
  grobl,
  which = c("column", "row"),
  space = unit(0.5, "mm"),
  width = NULL,
  height = NULL,
  gp = gpar(fill = NA, col = NA)
)
```

**Arguments**

grobl	A list of sequence logo grobs, typically created using <a href="#">seqLogoGrob</a> .
which	Whether it is a column annotation or a row annotation?
space	The space around the image to the annotation grid borders. The value should be a unit object.
width	Width of the annotation. The value should be an absolute unit. Width is not allowed to be set for column annotation.
height	Height of the annotation. The value should be an absolute unit. Height is not allowed to be set for row annotation.
gp	Graphic parameters for annotation grids. Can be used to control the background color in the annotation grids.

**Value**

An annotation function which can be used in [HeatmapAnnotation](#).

**Examples**

```
if (require(JASPAR2020) && require(TFBSTools) && require(gridExtra)) {
  pfm1 <- getMatrixByID(JASPAR2020, "MA0139")

  g1 <- seqLogoGrob(pfm1)

  anno <- annoSeqlogo(list(g1))
}
```

---

bin	<i>Bin elements of x.</i>
-----	---------------------------

---

### Description

bin groups elements of x into bins with either a constant number of elements per bin, a constant bin width or according to user-provided bin boundaries.

### Usage

```
bin(
  x,
  binmode = c("equalN", "equalWidth", "breaks"),
  nElements = round(length(x)/5),
  nBins = NULL,
  minAbsX = NULL,
  breaks = NULL,
  ...
)
```

### Arguments

x	A numerical vector with the values used for binning.
binmode	The algorithm to be used for binning. Possible values are: "equalN" (default), "equalWidth" or "breaks" (see Details).
nElements	The number of elements per bin (only for binmode="equalN"). The width of bins is adjusted accordingly.
nBins	The number of bins (only for binmode="equalWidth"). The number of elements per bin will be variable.
minAbsX	The minimal absolute value in x for elements to be binned using the binmode="equalN" or binmode="equalWidth" (ignored for other values of binmode). Elements with x values in $[-\text{minAbsX}, \text{minAbsX}]$ will be collected in a single bin.
breaks	Numerical vector with bin boundaries (only for binmode="breaks"). breaks has to be ordered and strictly increasing, and has to be of length (number of bins) + 1.
...	Further arguments to be passed to cut(x, breaks, include.lowest = TRUE, ...), such as labels=FALSE.

### Details

Elements are binned according to the values in x depending on binmode:

**equalN** Items are grouped into a variable number of bins with nElements elements each. If minAbsX is not NULL, elements with x-values in  $[-\text{minAbsX}, \text{minAbsX}]$  will first be collected in a single bin before binning the remaining elements. The boundaries of this single bin may be slightly adjusted in order to respect the nElements elements in the other bins.

**equalWidth** Items are group into nBins bins with a variable number of elements each.

**breaks** Items are grouped into bins using cut(x, breaks, include.lowest = TRUE)

**Value**

The return value from `cut(x, ...)`, typically a factor of the same length as `x`. Binning mode, bin boundaries and the "neutral" bin are available from `attr(..., "binmode")`, `attr(..., "breaks")` and `attr(..., "bin0")`. For `binmode = "breaks"`, the latter will be NA.

**See Also**

`cut` which is used internally.

**Examples**

```
set.seed(1)
x <- rnorm(100)
summary(bin(x, "equalN", nElements=10))
summary(bin(x, "equalN", nElements=10, minAbsX=0.5))
summary(bin(x, "equalWidth", nBins=5))
summary(bin(x, "breaks", breaks=c(-10,-1,0,1,10)))
```

---

calcBinnedKmerEnr

*Calculate k-mer enrichment in bins of sequences.*

---

**Description**

Given a set of sequences and corresponding bins, identify enriched k-mers (n-grams) in each bin. The sequences can be given either directly or as genomic coordinates.

**Usage**

```
calcBinnedKmerEnr(
  seqs,
  bins = NULL,
  kmerLen = 5,
  background = c("otherBins", "allBins", "zeroBin", "genome", "model"),
  MMorder = 1,
  test = c("fisher", "binomial"),
  includeRevComp = TRUE,
  maxFracN = 0.7,
  maxKmerSize = 3L,
  GCbreaks = c(0.2, 0.25, 0.3, 0.35, 0.4, 0.45, 0.5, 0.6, 0.7, 0.8),
  pseudocount.kmers = 1,
  pseudocount.log2enr = 8,
  p.adjust.method = "BH",
  genome = NULL,
  genome.regions = NULL,
  genome.oversample = 2,
  BPPARAM = SerialParam(),
  verbose = FALSE
)
```

**Arguments**

seqs	<a href="#">DNAStringSet</a> object with sequences to test
bins	Factor of the same length and order as seqs, indicating the bin for each sequence. Typically the return value of <a href="#">bin</a> . For background = "genome" or background = "model", bins can be omitted.
kmerLen	A numeric scalar giving the k-mer length.
background	A character scalar specifying the background sequences to use. One of "otherBins" (default), "allBins", "zeroBin", "genome" or "model" (see "Details").
MMorder	A numeric scalar giving the order of the Markov model used to calculate the expected frequencies for background = "model".
test	A character scalar specifying the type of enrichment test to perform. One of "fisher" (default) or "binomial". The enrichment test is one-sided (enriched in foreground).
includeRevComp	A logical scalar. If TRUE (default), count k-mer occurrences in both seqs and their reverse-complement, by concatenating seqs and their reverse-complemented versions before the counting. This is useful if motifs can be expected to occur on any strand (e.g. DNA sequences of ChIP-seq peaks). If motifs are only expected on the forward strand (e.g. RNA sequences of CLIP-seq peaks), includeRevComp = FALSE should be used. Note that bins will be recycled for the reverse complemented sequences, which means that each reverse-complemented sequence will be assigned to the same bin as the corresponding forward sequence.
maxFracN	A numeric scalar with the maximal fraction of N bases allowed in a sequence (defaults to 0.7). Sequences with higher fractions are excluded from the analysis.
maxKmerSize	The maximum k-mer size to consider, when adjusting background sequence weights for k-mer composition compared to the foreground sequences. The default value (3) will correct for mono-, di- and tri-mer composition.
GCbreaks	The breaks between GC bins. The default value is based on the hard-coded bins used in Homer.
pseudocount.kmers	A numeric scalar - will be added to the observed and expected counts for each k-mer to avoid zero values.
pseudocount.log2enr	A numerical scalar with the pseudocount to add to foreground and background counts when calculating log2 motif enrichments
p.adjust.method	A character scalar selecting the p value adjustment method (used in <a href="#">p.adjust</a> ).
genome	A BSgenome or DNAStringSet object with the genome sequence. Only used for background = "genome" for extracting background sequences.
genome.regions	An optional <a href="#">GRanges</a> object defining the intervals in genome from which background sequences are sampled for background = "genome". If NULL, background sequences are sampled randomly from genome.
genome.oversample	A numeric scalar of at least 1.0 defining how many background sequences will be sampled per foreground sequence for background = "genome". Larger values will take longer but improve the sequence composition similarity between foreground and background (see "Details").

BPPARAM	An optional <a href="#">BiocParallelParam</a> instance determining the parallel back-end to be used during evaluation.
verbose	A logical scalar. If TRUE, report on progress.

## Details

This function implements a binned k-mer enrichment analysis. In each enrichment analysis, the sequences in a specific bin are used as foreground sequences to test for k-mer enrichments comparing to background sequences (defined by background, see below), similarly as in done for motifs in [calcBinnedMotifEnrR](#). Sequences are weighted to correct for GC and shorter k-mer composition differences between fore- and background sets.

The background sequences are defined according to the value of the background argument:

**otherBins** : sequences from all other bins (excluding the current bin)

**allBins** : sequences from all bins (including the current bin)

**zeroBin** : sequences from the "zero bin", defined by the maxAbsX argument of [bin](#). If bins does not define a "zero bin", for example because it was created by `bin(..., maxAbsX = NULL)`, selecting this background definition will abort with an error.

**genome** : sequences randomly sampled from the genome (or the intervals defined in `genome.regions` if given). For each foreground sequence, `genome.oversample` background sequences of the same size are sampled (on average). From these, one per foreground sequence is selected trying to match the G+C composition. In order to make the sampling deterministic, a seed number needs to be provided to the RNGseed parameter in [SerialParam](#) or [MulticoreParam](#) when creating the [BiocParallelParam](#) instance in BPPARAM.

**model** : a Markov model of the order MMorder is estimated from the foreground sequences and used to estimate expected k-mer frequencies. K-mer enrichments are then calculated comparing observed to these expected frequencies. In order to make the process deterministic, a seed number needs to be provided to the RNGseed parameter in [SerialParam](#) or [MulticoreParam](#) when creating the [BiocParallelParam](#) instance in BPPARAM.

For each k-mer, the weights of sequences is multiplied with the number of k-mer occurrences in each sequence and summed, separately for foreground (`sumForegroundWgtWithHits`) and background (`sumBackgroundWgtWithHits`) sequences. The function works in ZOOPS (Zero-Or-One-Per-Sequence) mode, so at most one occurrence per sequence is counted, which helps reduce the impact of sequence repeats. The total foreground (`totalWgtForeground`) and background (`totalWgtBackground`) sum of sequence weights is also calculated. If a k-mer has zero `sumForegroundWgtWithHits` and `sumBackgroundWgtWithHits`, then any values (p-values and enrichment) that are calculated using these two numbers are set to NA.

Two statistical tests for the calculation of enrichment log p-value are available: `test = "fisher"` (default) to perform Fisher's exact tests, or `test = "binomial"` to perform binomial tests, using:

**fisher** : `fisher.test(x = tab, alternative = "greater")`, where `tab` is the contingency table with the summed weights of sequences in foreground or background sets (rows), and with or without a occurrences of a particular k-mer (columns).

**binomial** : `pbinom(q = sumForegroundWgtWithHits - 1, size = totalWgtForeground, prob = sumBackgroundWgtWithHits / totalWgtBackground, lower.tail = FALSE, log.p = TRUE)`

## Value

A [SummarizedExperiment](#) object with motifs in rows and bins in columns, containing seven assays:

**negLog10P** : -log10 P values



**negLog10Padj** : -log10 adjusted P values

**pearsonResid** : k-mer enrichments as Pearson residuals

**expForegroundWgtWithHits** : expected number of foreground sequences with motif hits

**log2enr** : k-mer enrichments as log2 ratios

**sumForegroundWgtWithHits** : Sum of foreground sequence weights in a bin that have k-mer occurrences

**sumBackgroundWgtWithHits** : Sum of background sequence weights in a bin that have k-mer occurrences

#' The rowData of the object contains annotations (name, PFMs, PWMs and GC fraction) for the k-mers, while the colData slot contains summary information about the bins.

### See Also

[getKmerFreq](#) used to calculate k-mer enrichments; [getSeq,BSgenome-method](#) which is used to extract sequences from genomepkg if x is a GRanges object; [bplapply](#) that is used for parallelization; [bin](#) for binning of regions

### Examples

```
seqs <- Biostrings::DNAStringSet(c("GCATGCATGC", "CATGCATGC"))
bins <- factor(1:2)
calcBinnedKmerEnr(seqs = seqs, bins = bins, kmerLen = 3)
```

---

calcBinnedMotifEnrHomer

*Prepare and run HOMER motif enrichment analysis.*

---

### Description

Run complete HOMER motif enrichment analysis, consisting of calls to [prepareHomer](#), [system2](#) and [parseHomerOutput](#). This function requires HOMER to be installed (see <http://homer.ucsd.edu/homer/index.html>) and the path to the tool to be provided (homerfile argument).

### Usage

```
calcBinnedMotifEnrHomer(
  gr,
  b,
  genomedir,
  outdir,
  motifFile,
  homerfile = findHomer(),
  regionsize = "given",
  pseudocount.log2enr = 8,
  p.adjust.method = "BH",
  Ncpu = 2L,
  verbose = FALSE,
  verbose.Homer = FALSE
)
```

## Arguments

<code>gr</code>	A GRanges object (or an object that can be coerced to one) with the genomic regions to analyze.
<code>b</code>	A vector of the same length as <code>gr</code> that groups its elements into bins (typically a factor, such as the one returned by <a href="#">bin</a> ).
<code>genomedir</code>	Directory containing sequence files in Fasta format (one per chromosome).
<code>outdir</code>	A path specifying the folder into which the output files will be written.
<code>motifFile</code>	A file with HOMER formatted PWMs to be used in the enrichment analysis.
<code>homerfile</code>	Path and file name of the <code>findMotifsGenome.pl</code> HOMER script.
<code>regionsize</code>	The peak size to use in HOMER ("given" keeps the coordinate region, an integer value will keep only that many bases in the region center).
<code>pseudocount.log2enr</code>	A numerical scalar with the pseudocount to add to foreground and background counts when calculating log2 motif enrichments
<code>p.adjust.method</code>	A character scalar selecting the p value adjustment method (used in <a href="#">p.adjust</a> ).
<code>Ncpu</code>	Number of parallel threads that HOMER can use.
<code>verbose</code>	A logical scalar. If TRUE, print progress messages.
<code>verbose.Homer</code>	A logical scalar. If TRUE, print the console output when running Homer.

## Value

A SummarizedExperiment object with motifs in rows and bins in columns, containing seven assays:

**negLog10P** : -log10 P values

**negLog10Padj** : -log10 adjusted P values

**pearsonResid** : motif enrichments as Pearson residuals

**expForegroundWgtWithHits** : expected number of foreground sequences with motif hits

**log2enr** : motif enrichments as log2 ratios

**sumForegroundWgtWithHits** : Sum of foreground sequence weights in a bin that have motif hits

**sumBackgroundWgtWithHits** : Sum of background sequence weights in a bin that have motif hits

The `rowData` of the object contains annotations (name, PFMs, PWMs and GC fraction) for the motifs, while the `colData` slot contains summary information about the bins.

## See Also

The functions that are wrapped: [prepareHomer](#), [system2](#) and [parseHomerOutput](#), [bin](#) for binning of regions

## Examples

```
if (!is.na(findHomer())){

  # genome
  genome <- system.file("extdata", "exampleGenome.fa", package = "monaLisa")

  # create motif file for Homer
```

```

motiffile <- tempfile()
motifIDs <- c("MA0139.1", "MA1102.1", "MA0740.1")
dumpJaspar(filename = motiffile, pkg = "JASPAR2020",
            opts = list(ID = motifIDs))

# GRanges of regions used in binned motif enrichment analysis
gr <- GenomicRanges::tileGenome(
  seqlengths = c(chr1 = 10000L, chr2 = 10000L, chr3 = 10000L),
  tilewidth = 200, cut.last.tile.in.chrom = TRUE)

# create bins (motif enrichment analysis will be per bin)
bins <- factor(GenomicRanges::seqnames(gr))
table(bins)

# run calcBinnedMotifEnrHomer
outdir <- tempfile()
se <- calcBinnedMotifEnrHomer(gr = gr, b = bins, genomedir = genome,
  outdir = outdir, motifFile = motiffile)
list.files(outdir)

}

```

---

calcBinnedMotifEnrR      *Binned Motif Enrichment Analysis with monalisa*

---

## Description

This function performs a motif enrichment analysis on bins of sequences. For each bin, the sequences in all other bins are used as background.

## Usage

```

calcBinnedMotifEnrR(
  seqs,
  bins = NULL,
  pwmL = NULL,
  background = c("otherBins", "allBins", "zeroBin", "genome"),
  test = c("fisher", "binomial"),
  maxFracN = 0.7,
  maxKmerSize = 3L,
  min.score = 10,
  matchMethod = "matchPWM",
  GCbreaks = c(0.2, 0.25, 0.3, 0.35, 0.4, 0.45, 0.5, 0.6, 0.7, 0.8),
  pseudocount.log2enr = 8,
  p.adjust.method = "BH",
  genome = NULL,
  genome.regions = NULL,
  genome.oversample = 2,
  BPPARAM = SerialParam(),
  verbose = FALSE,
  ...
)

```

**Arguments**

seqs	<a href="#">DNAStringSet</a> object with sequences to test
bins	Factor of the same length and order as seqs, indicating the bin for each sequence. Typically the return value of <a href="#">bin</a> . For background = "genome", bins can be omitted.
pwmL	PWMMatrixList with motifs for which to calculate enrichments.
background	A character scalar specifying the background sequences to use. One of "otherBins" (default), "allBins", "zeroBin" or "genome" (see "Details").
test	A character scalar specifying the type of enrichment test to perform. One of "fisher" (default) or "binomial". The enrichment test is one-sided (enriched in foreground).
maxFracN	A numeric scalar with the maximal fraction of N bases allowed in a sequence (defaults to 0.7). Sequences with higher fractions are excluded from the analysis.
maxKmerSize	The maximum k-mer size to consider, when adjusting background sequence weights for k-mer composition compared to the foreground sequences. The default value (3) will correct for mono-, di- and tri-mer composition.
min.score	The minimal score for motif hits, used in <a href="#">findMotifHits</a> .
matchMethod	The method used to scan for motif hits, passed to the method parameter in <a href="#">findMotifHits</a> .
GCbreaks	The breaks between GC bins. The default value is based on the hard-coded bins used in Homer.
pseudocount.log2enr	A numerical scalar with the pseudocount to add to foreground and background counts when calculating log2 motif enrichments
p.adjust.method	A character scalar selecting the p value adjustment method (used in <a href="#">p.adjust</a> ).
genome	A BSgenome or DNAStringSet object with the genome sequence. Only used for background = "genome" for extracting background sequences.
genome.regions	An optional <a href="#">GRanges</a> object defining the intervals in genome from which background sequences are sampled for background = "genome". If NULL, background sequences are sampled randomly from genome.
genome.oversample	A numeric scalar of at least 1.0 defining how many background sequences will be sampled per foreground sequence for background = "genome". Larger values will take longer but improve the sequence composition similarity between foreground and background (see "Details").
BPPARAM	An optional <a href="#">BiocParallelParam</a> instance determining the parallel back-end to be used during evaluation.
verbose	A logical scalar. If TRUE, print progress messages.
...	Additional arguments for <a href="#">findMotifHits</a> .

**Details**

This function implements a binned motif enrichment analysis. In each enrichment analysis, the sequences in a specific bin are used as foreground sequences to test for motif enrichments comparing to background sequences (defined by background, see below). The logic follows the `findMotifsGenome.pl`

tool from Homer version 4.11, with `-size` given `-nomotif -mknown` and additionally `-h` if using `test = "fisher"`, and gives very similar results. As in the Homer tool, sequences are weighted to correct for GC and k-mer composition differences between fore- and background sets.

The background sequences are defined according to the value of the background argument:

**otherBins** : sequences from all other bins (excluding the current bin)

**allBins** : sequences from all bins (including the current bin)

**zeroBin** : sequences from the "zero bin", defined by the `maxAbsX` argument of `bin`. If bins does not define a "zero bin", for example because it was created by `bin(..., maxAbsX = NULL)`, selecting this background definition will abort with an error.

**genome** : sequences randomly sampled from the genome (or the intervals defined in `genome.regions` if given). For each foreground sequence, `genome.oversample` background sequences of the same size are sampled (on average). From these, one per foreground sequence is selected trying to match the G+C composition. In order to make the sampling deterministic, a seed number needs to be provided to the `RNGseed` parameter in `SerialParam` or `MulticoreParam` when creating the `BiocParallelParam` instance in `BPPARAM`.

Motif hits are predicted using `findMotifHits` and multiple hits per sequence are counted as just one hit (ZOOPS mode). For each motif, the weights of sequences that have a hit are summed separately for foreground (`sumForegroundWgtWithHits`) and background (`sumBackgroundWgtWithHits`). The total foreground (`totalWgtForeground`) and background (`totalWgtBackground`) sum of sequence weights is also calculated. If a motif has zero `sumForegroundWgtWithHits` and `sumBackgroundWgtWithHits`, then any values (p-values and enrichment) that are calculated using these two numbers are set to NA.

Two statistical tests for the calculation of enrichment log p-value are available: `test = "fisher"` (default) to perform Fisher's exact tests, or `test = "binomial"` to perform binomial tests (default in Homer), using:

**fisher** : `fisher.test(x = tab, alternative = "greater")`, where `tab` is the contingency table with the summed weights of sequences in foreground or background sets (rows), and with or without a hit for a particular motif (columns).

**binomial** : `pbinom(q = sumForegroundWgtWithHits - 1, size = totalWgtForeground, prob = sumBackgroundWgtWithHits / totalWgtBackground, lower.tail = FALSE, log.p = TRUE)`

## Value

A `SummarizedExperiment` object with motifs in rows and bins in columns, containing seven assays:

**negLog10P** :  $-\log_{10}$  P values

**negLog10Padj** :  $-\log_{10}$  adjusted P values

**pearsonResid** : motif enrichments as Pearson residuals

**expForegroundWgtWithHits** : expected number of foreground sequences with motif hits

**log2enr** : motif enrichments as  $\log_2$  ratios

**sumForegroundWgtWithHits** : Sum of foreground sequence weights in a bin that have motif hits

**sumBackgroundWgtWithHits** : Sum of background sequence weights in a bin that have motif hits

The `rowData` of the object contains annotations (name, PFMs, PWMs and GC fraction) for the motifs, while the `colData` slot contains summary information about the bins.

**Examples**

```
seqs <- Biostrings::DNAStringSet(c("GTCAGTCGATC", "CAGTCTAGCTG",
                                   "CGATCGTCAGT", "AGCTGCAGTCT"))

bins <- factor(rep(1:2, each = 2))

m <- rbind(A = c(2, 0, 0),
          C = c(1, 1, 0),
          G = c(0, 2, 0),
          T = c(0, 0, 3))

pwms <- TFBSTools::PWMMatrixList(
  TFBSTools::PWMMatrix(ID = "m1", profileMatrix = m),
  TFBSTools::PWMMatrix(ID = "m2", profileMatrix = m[, 3:1])
)

calcBinnedMotifEnrR(seqs = seqs, bins = bins, pwml = pwms,
                    min.score = 3)
```

dumpJaspar

*Dump Jaspar motifs into a HOMER motif file.***Description**

Get motifs from a Jaspar database package (e.g. JASPAR2020) and write them into a HOMER-compatible motif file as positional probability matrices.

**Usage**

```
dumpJaspar(
  filename,
  pkg = "JASPAR2020",
  opts = list(tax_group = "vertebrates"),
  pseudocount = 1,
  relScoreCutoff = 0.8,
  verbose = FALSE
)
```

**Arguments**

filename	Name of the output file to be created.
pkg	Name of the Jaspar package to use (default: JASPAR2020).
opts	A list with search options used in <a href="#">getMatrixSet</a> . By default, only vertebrate motifs are included in the output using <code>opts = list(tax_group = "vertebrates")</code> .
pseudocount	A numerical scalar with the pseudocount to be added to each element of the position frequency matrix extracted from Jaspar, before its conversion to a position probability matrix (default: 1.0).
relScoreCutoff	Currently ignored. numeric(1) in [0,1] that sets the default motif log-odds score cutoff to <code>relScoreCutoff * maximal score for each PWM</code> (default: 0.8).
verbose	A logical scalar. If TRUE, print progress messages.

**Value**

TRUE if successful.

## See Also

[getMatrixSet](#) for details on the argument `opts`. [homerToPFMatrixList](#) to read a file with HOMER-formatted motifs into a [PFMatrixList](#).

## Examples

```
dumpJaspar(filename = tempfile(), pkg = "JASPAR2020",
            opts = list(ID = c("MA0006.1")))
```

---

findHomer	<i>Find HOMER script file.</i>
-----------	--------------------------------

---

## Description

Find absolute path to HOMER script file.

## Usage

```
findHomer(homerfile = "findMotifsGenome.pl", dirs = NULL)
```

## Arguments

<code>homerfile</code>	Name of the script file to search.
<code>dirs</code>	Directory names to look for <code>homerfile</code> . If <code>dirs=NULL</code> , all directories listed in the <code>PATH</code> environment variable will be searched.

## Details

In addition to `dirs`, `findHomer` will also look in the directory provided in the environment variable `MONALISA_HOMER`.

## Value

Absolute path to `homerfile`, or `NA` if none or several were found.

## Examples

```
homer_path <- findHomer()
```

---

findMotifHits*Find motif matches in sequences.*

---

**Description**

findMotifHits scans sequences (either provided as a file, an R object or genomic coordinates) for matches to positional weight matrices (provided as a file or as R objects)

**Usage**

```
findMotifHits(  
  query,  
  subject,  
  min.score,  
  method = c("matchPWM", "homer2"),  
  homerfile = findHomer("homer2"),  
  BPPARAM = SerialParam(),  
  genome = NULL  
)  
  
## S4 method for signature 'character,character'  
findMotifHits(  
  query,  
  subject,  
  min.score,  
  method = c("matchPWM", "homer2"),  
  homerfile = findHomer("homer2"),  
  BPPARAM = SerialParam(),  
  genome = NULL  
)  
  
## S4 method for signature 'character,DNAString'  
findMotifHits(  
  query,  
  subject,  
  min.score,  
  method = c("matchPWM", "homer2"),  
  homerfile = findHomer("homer2"),  
  BPPARAM = SerialParam(),  
  genome = NULL  
)  
  
## S4 method for signature 'character,DNAStringSet'  
findMotifHits(  
  query,  
  subject,  
  min.score,  
  method = c("matchPWM", "homer2"),  
  homerfile = findHomer("homer2"),  
  BPPARAM = SerialParam(),  
  genome = NULL  
)
```



```
)

## S4 method for signature 'PWMMatrix,character'
findMotifHits(
  query,
  subject,
  min.score,
  method = c("matchPWM", "homer2"),
  homerfile = findHomer("homer2"),
  BPPARAM = SerialParam(),
  genome = NULL
)

## S4 method for signature 'PWMMatrix,DNAString'
findMotifHits(
  query,
  subject,
  min.score,
  method = c("matchPWM", "homer2"),
  homerfile = findHomer("homer2"),
  BPPARAM = SerialParam(),
  genome = NULL
)

## S4 method for signature 'PWMMatrix,DNAStringSet'
findMotifHits(
  query,
  subject,
  min.score,
  method = c("matchPWM", "homer2"),
  homerfile = findHomer("homer2"),
  BPPARAM = SerialParam(),
  genome = NULL
)

## S4 method for signature 'PWMMatrixList,character'
findMotifHits(
  query,
  subject,
  min.score,
  method = c("matchPWM", "homer2"),
  homerfile = findHomer("homer2"),
  BPPARAM = SerialParam(),
  genome = NULL
)

## S4 method for signature 'PWMMatrixList,DNAString'
findMotifHits(
  query,
  subject,
  min.score,
  method = c("matchPWM", "homer2"),
```

```

    homerfile = findHomer("homer2"),
    BPPARAM = SerialParam(),
    genome = NULL
)

## S4 method for signature 'PWMMatrixList,DNAStringSet'
findMotifHits(
  query,
  subject,
  min.score,
  method = c("matchPWM", "homer2"),
  homerfile = findHomer("homer2"),
  BPPARAM = SerialParam(),
  genome = NULL
)

## S4 method for signature 'PWMMatrix,GRanges'
findMotifHits(
  query,
  subject,
  min.score,
  method = c("matchPWM", "homer2"),
  homerfile = findHomer("homer2"),
  BPPARAM = SerialParam(),
  genome = NULL
)

## S4 method for signature 'PWMMatrixList,GRanges'
findMotifHits(
  query,
  subject,
  min.score,
  method = c("matchPWM", "homer2"),
  homerfile = findHomer("homer2"),
  BPPARAM = SerialParam(),
  genome = NULL
)

```

### Arguments

query	<p>The motifs to search for, either a  character(1) with the path and file name of a motif file with PWM in HOMER  format (currently only supported for method="homer2")  PWMMatrix with a single PWM  PWMMatrixList with several PWMs to search for.</p>
subject	<p>The sequences to be searched, either a  character with the path and file name of a sequence file with DNA sequences  in FASTA format  DNAString with a single sequence  DNAStringSet with several sequences  GRanges object with the genomic coordinates of the sequences to be searched.</p>

min.score	The minimum score for counting a match. Can be given as a character string containing a percentage (e.g. "85 highest possible score or as a single number.
method	The internal method to use for motif searching. One of "matchPWM" using Biostrings::matchPWM (optimized) "homer2" call to the homer2 binary Please note that the two methods might give slightly different results (see details).
homerfile	Path and file name of the homer2 binary.
BPPARAM	An optional <a href="#">BiocParallelParam</a> instance determining the parallel back-end to be used during evaluation.
genome	BSgenome object that is the reference genome of the subject. This argument is set to NULL by default and only used by the function when the subject is a GRanges object. It is then necessary to specify the genome so that the function can internally convert the genomic regions into a DNASTringSet object.

## Details

The implemented methods (matchPWM and homer2) are there for convenience (method="matchPWM" calls Biostrings::matchPWM internally in an optimized fashion, and method = "homer2" calls the command line tool from Homer and therefore requires an installation of Homer).

In general, running findMotifHits with the same parameters using any of the methods generates identical results. Some minor differences could occur that result from rounding errors during the necessary conversion of PWMs (log2-odd scores) to the probability matrices needed by Homer, and the conversion of scores from and to the natural log scale used by Homer. These conversions are implemented transparently for the user, so that the arguments of findMotifHits do not have to be adjusted (e.g. the PWMs should always contain log2-odd scores, and min.score is always on the log2 scale).

If there are bases with frequencies of less than 0.001 in a motif, Homer will set them to 0.001 and adjust the other frequencies at that motif position accordingly so that they sum to 1.0. This may differ from the adjustment used when scanning a PWM with matchPWM (e.g. the pseudocounts argument in the [toPWM](#) function), and thus can give rise to differences in reported motif hits and hit scores (typically only low-scoring hits).

## Value

A GRanges object with the matches to query in subject.

## Examples

```
seqs <- Biostrings::DNASTringSet(c(s1 = "GTCAGTCGATC", s2 = "CAGTCTAGCTG",
                                   s3 = "CGATCGTCAGT", s4 = "AGCTGCACTCT"))
m <- rbind(A = c(2, 0, 0),
           C = c(1, 1, 0),
           G = c(0, 2, 0),
           T = c(0, 0, 3))
pwms <- TFBSTools::PWMMatrixList(
  TFBSTools::PWMMatrix(ID = "m1", profileMatrix = m),
  TFBSTools::PWMMatrix(ID = "m2", profileMatrix = m[, 3:1])
)
findMotifHits(pwms, seqs, min.score = 7)
```

---

getColsByBin	<i>Get colors by bin.</i>
--------------	---------------------------

---

## Description

Get colors for elements according to their bin. Colors are assigned to bins forming a gradient from col1 to col2 in the order of levels{b}. col0 is assigned to the neutral bin (attribute "") if available.

## Usage

```
getColsByBin(  
  b,  
  col1 = c("#003C30", "#01665E", "#35978F", "#80CDC1", "#C7EAE5"),  
  col2 = c("#F6E8C3", "#DFC27D", "#BF812D", "#8C510A", "#543005"),  
  col0 = "#F5F5F5"  
)
```

## Arguments

b	A factor that groups elements into bins (typically the output of <a href="#">bin</a> ).
col1	First color.
col2	Second color.
col0	Neutral color.

## Value

A character vector with colors for the elements in b.

## See Also

[bin](#).

## Examples

```
set.seed(1)  
x <- rnorm(100)  
b <- bin(x, "equalN", nElements = 10)  
cols <- getColsByBin(b)
```

getKmerFreq

*Calculate observed and expected k-mer frequencies***Description**

Given a set of sequences, calculate observed and expected k-mer frequencies. Expected frequencies are based on a Markov model of order MMorder.

**Usage**

```
getKmerFreq(
  seqs,
  kmerLen = 5,
  MMorder = 1,
  pseudocount = 1,
  zoops = TRUE,
  strata = rep(1L, length(seqs)),
  p.adjust.method = "BH",
  includeRevComp = TRUE
)
```

**Arguments**

seqs	Set of sequences, either a character vector or a <a href="#">DNAStringSet</a> .
kmerLen	A numeric scalar giving the k-mer length.
MMorder	A numeric scalar giving the order of the Markov model used to calculate the expected frequencies.
pseudocount	A numeric scalar - will be added to the observed counts for each k-mer to avoid zero values.
zoops	A logical scalar. If TRUE (the default), only one or zero occurrences of a k-mer are considered per sequence.
strata	A factor or a numeric scalar defining the strata of sequences. A separate Markov model and expected k-mer frequencies are estimated for the set of sequences in each stratum (level in a strata factor). If strata is a scalar value, it will be interpreted as the number of strata to split the sequences into according to their CpG observed-over-expected counts using <code>kmeans(CpGoe, centers = strata)</code> .
p.adjust.method	A character scalar selecting the p value adjustment method (used in <a href="#">p.adjust</a> ).
includeRevComp	A logical scalar. If TRUE (default), count k-mer occurrences in both seqs and their reverse-complement, by concatenating seqs and their reverse-complemented versions before the counting. This is useful if motifs can be expected to occur on any strand (e.g. DNA sequences of ChIP-seq peaks). If motifs are only expected on the forward strand (e.g. RNA sequences of CLIP-seq peaks), <code>includeRevComp = FALSE</code> should be used. Note that if strata is a vector of the same length as seqs, each reverse-complemented sequence will be assigned to the same stratum as the forward sequence.

**Value**

A list with observed and expected k-mer frequencies (`freq.obs` and `freq.exp`, respectively), and enrichment statistics for each k-mer.

**Examples**

```
res <- getKmerFreq(seqs = c("AAAAATT", "AAATTTT"), kmerLen = 3)
names(res)
head(res$freq.obs)
head(res$freq.exp)
```

---

getSetZeroBin	<i>Get and set the zero bin manually</i>
---------------	--

---

**Description**

Get and set the zero bin manually

**Usage**

```
getZeroBin(bins)

setZeroBin(bins, zeroBin)
```

**Arguments**

<code>bins</code>	Factor, typically the return value of <a href="#">bin</a> .
<code>zeroBin</code>	Numeric or character scalar indicating the level to use as the zero bin, or NA.

**Value**

For `getZeroBin`, the index of the level representing the zero bin. For `setZeroBin`, a modified factor with the zero bin set to the provided value.

**Examples**

```
set.seed(1)
x <- rnorm(100)
bins <- bin(x, "equalN", nElements = 10, minAbsX = 0.5)
getZeroBin(bins)
bins <- setZeroBin(bins, 2)
```

---

homerToPFMatrixList      *Read a HOMER motif file and create a PFMatrixList*

---

### Description

Read motifs from a file in HOMER format and create a PFMatrixList from them.

### Usage

```
homerToPFMatrixList(filename, n = 100L)
```

### Arguments

filename	Name of the input file with HOMER-formatted motifs.
n	The number of observations (multiplied with base frequencies to create the number of observed bases at each position).

### Value

A [PFMatrixList](#) with motifs from the file.

### See Also

[dumpJaspar](#) for writing motifs from a Jaspar database package into a file in HOMER format.

### Examples

```
library(JASPAR2020)
optsL <- list(ID = c("MA0006.1"))
pfm1 <- TFBSTools::getMatrixSet(JASPAR2020, opts = optsL)
TFBSTools::Matrix(pfm1)

tmpfn <- tempfile()
dumpJaspar(filename = tmpfn, pkg = "JASPAR2020", opts = optsL)
pfm2 <- homerToPFMatrixList(tmpfn)
TFBSTools::Matrix(pfm2)

unlink(tmpfn)
```

---

motifKmerSimilarity      *Calculate similarities between motifs and k-mers.*

---

### Description

For each motif, calculate its similarity to all k-mers of length kmerLen, defined as the maximal probability of observing the k-mer given the base frequencies of the motif (the maximum is taken over for all possible ungapped alignments between motif and k-mer). If necessary matrices are padded on the sides with background base frequencies (assuming all bases to have a frequency of 0.25).

**Usage**

```
motifKmerSimilarity(
  x,
  kmerLen = 5,
  kmers = NULL,
  includeRevComp = FALSE,
  BPPARAM = SerialParam(),
  verbose = FALSE
)
```

**Arguments**

x	Either a <a href="#">PFMatrixList</a> , or a character scalar with a file containing motifs in HOMER format (used directly method = "HOMER", loaded into a <a href="#">PFMatrixList</a> by <a href="#">homerToPFMatrixList</a> for method = "R").
kmerLen	A numeric scalar giving the k-mer length.
kmers	Either a character vector of k-mers for which to calculate the similarity to each motif, or NULL, in which case all k-mers of length kmerLen are used.
includeRevComp	A logical scalar. If set to TRUE, each k-mer as well as its reverse complement is compared to each motif, and the larger of the two similarities is returned.
BPPARAM	An optional <a href="#">BiocParallelParam</a> instance determining the parallel back-end to be used during evaluation.
verbose	A logical scalar. If TRUE, report on progress.

**Value**

A matrix of probabilities for each motif - k-mer pair.

**See Also**

[bplapply](#) used for parallelization.

**Examples**

```
m <- rbind(A = c(12, 0, 0),
           C = c( 3, 2, 0),
           G = c( 0, 14, 0),
           T = c( 0, 0, 15))
pfms <- TFBSTools::PFMatrixList(
  TFBSTools::PFMatrix(name = "m1", profileMatrix = m),
  TFBSTools::PFMatrix(name = "m2", profileMatrix = m[, 3:1])
)
motifKmerSimilarity(pfms, kmerLen = 3)[, c("AGT", "TGA")]
```



motifSimilarity

*Calculate similarities between pairs of motifs.***Description**

For each pair of motifs, calculate the similarity defined as the maximal Pearson's correlation coefficient between base frequencies over all possible shifts (relative positions of the two matrices with at least one overlapping position). If necessary matrices are padded on the sides with background base frequencies (assuming all bases to have a frequency of 0.25) to enable comparison of all positions in both matrices.

**Usage**

```
motifSimilarity(
  x,
  y = NULL,
  method = c("R", "HOMER"),
  homerfile = findHomer("compareMotifs.pl"),
  homerOutfile = NULL,
  BPPARAM = SerialParam(),
  verbose = FALSE
)
```

**Arguments**

x	Either a <a href="#">PFMatrixList</a> , or a character scalar with a file containing motifs in HOMER format (used directly method = "HOMER", loaded into a <a href="#">PFMatrixList</a> by <a href="#">homerToPFMatrixList</a> for method = "R").
y	Either a <a href="#">PFMatrixList</a> or NULL (default). If y = NULL, then similarities will be calculated for all pairs of motifs within x. Otherwise, method must be "R" and similarities will be calculated between any motif from x to any motif from y.
method	A character scalar specifying the method for similarity calculations. Either "R" (pure R implementation) or "HOMER" (will call the compareMotifs.pl script from HOMER). Results are identical (apart from rounding errors), and the R implementation is usually faster and can be parallelized (BPPARAM argument).
homerfile	Path to the HOMER script compareMotifs.pl (only used for method = "HOMER").
homerOutfile	A character scalar giving the file to save the similarity scores (only for method = "HOMER"). If NULL, scores will be stored into a temporary file.
BPPARAM	An optional <a href="#">BiocParallelParam</a> instance determining the parallel back-end to be used during evaluation (only used for method = "R").
verbose	A logical scalar. If TRUE, report on progress.

**Value**

A matrix of Pearson's correlation coefficients for each pair of motifs.

**See Also**

[bplapply](#) used for parallelization for method = "R", documentation of HOMER's compareMotifs.pl for details on method = "HOMER".

**Examples**

```

m <- rbind(A = c(12, 0, 0),
           C = c( 3, 2, 0),
           G = c( 0, 14, 0),
           T = c( 0, 0, 15))
pfms <- TFBSTools::PFMatrixList(
  TFBSTools::PFMatrix(name = "m1", profileMatrix = m),
  TFBSTools::PFMatrix(name = "m2", profileMatrix = m + 10),
  TFBSTools::PFMatrix(name = "m3", profileMatrix = m[, 3:1])
)
motifSimilarity(pfms)

```

---

 parseHomerOutput

*Load output from HOMER findMotifsGenome.pl into R*


---

**Description**

Parse HOMER output files into R data structures.

**Usage**

```
parseHomerOutput(infiles, pseudocount.log2enr = 8, p.adjust.method = "BH")
```

**Arguments**

**infiles** HOMER output files to be parsed.

**pseudocount.log2enr** A numerical scalar with the pseudocount to add to foreground and background counts when calculating log2 motif enrichments

**p.adjust.method** A character scalar selecting the p value adjustment method (used in [p.adjust](#)).

**Value**

A list of nine components (negLog10P, negLog10Padj, pearsonResid, expForegroundWgtWithHits, log2enr, sumForegroundWgtWithHits and sumBackgroundWgtWithHits), seven containing each a motif (rows) by bin (columns) matrix with raw -log10 P values, -log10 adjusted P values, the expected number of foreground sequences with hits, the observed number of foreground and background sequences with hits, and motif enrichments as Pearson residuals (pearsonResid) and as log2 ratios (log2enr), and two containing the total foreground and background weight (totalWgtForeground, totalWgtBackground).

**Examples**

```

outfile <- system.file("extdata", "homer_output.txt.gz",
                      package = "monaLisa")
res <- parseHomerOutput(infiles = c(bin1 = outfile))
head(res$negLog10P)

```

---

plotBinDensity	<i>Density plot of binned elements.</i>
----------------	---

---

## Description

Plot the density of binned elements with binning information.

## Usage

```
plotBinDensity(
  x,
  b,
  xlab = deparse(substitute(x, env = as.environment(-1))),
  ylab = "Density",
  main = "",
  legendPosition = "right",
  legend = NULL,
  legend.cex = NULL,
  ...
)
```

## Arguments

<code>x</code>	A numerical vector with the values used for binning.
<code>b</code>	A factor that groups elements of <code>x</code> into bins (typically the output of <a href="#">bin</a> ).
<code>xlab</code> , <code>ylab</code> , <code>main</code>	character scalars that set the x-axis label, y-axis label and the main title. Use "" to suppress the label.
<code>legendPosition</code>	A character scalar. If not "none", draw a legend with binning information. The value is used to control the legend position and will be passed to <code>theme(legend.position = legendPosition)</code> .
<code>legend</code>	Deprecated (ignored). Please use <code>legendPosition</code> to control the drawing and position of the legend.
<code>legend.cex</code>	Deprecated (ignored). You can use <a href="#">theme</a> to set legend and other graphical parameters.
<code>...</code>	Further arguments passed to <a href="#">getColsByBin</a> .

## Value

The generated density plot as a ggplot object.

## See Also

[getColsByBin](#), [geom\\_density](#)

## Examples

```
set.seed(1)
x <- rnorm(100)
b <- bin(x, "equalN", nElements = 10)
plotBinDensity(x, b)
```

---

plotBinDiagnostics	<i>Plot diagnostics of binned sequences</i>
--------------------	---

---

## Description

Plot various diagnostics of binned sequences. Three plot types are available:

`length` plots the distribution of sequence lengths within each bin.

`GCfrac` plots the distribution of GC fractions within each bin.

`dinucfreq` plots a heatmap of the relative frequency of each dinucleotide, averaged across the sequences within each bin. The values are centered for each dinucleotide to better highlight differences between the bins. The average relative frequency of each dinucleotide (across the bins) is indicated as well.

## Usage

```
plotBinDiagnostics(
  seqs,
  bins,
  aspect = c("length", "GCfrac", "dinucfreq"),
  draw_quantiles = c(0.25, 0.5, 0.75),
  ...
)
```

## Arguments

<code>seqs</code>	<a href="#">DNASTringSet</a> object with sequences.
<code>bins</code>	Factor of the same length and order as <code>seqs</code> , indicating the bin for each sequence. Typically the return value of <code>bin</code> .
<code>aspect</code>	The diagnostic to plot. Should be one of "length", "GCfrac" and "dinucfreq", to plot the distribution of sequence lengths, the distribution of GC fractions and the average relative dinucleotide frequencies across the bins.
<code>draw_quantiles</code>	For <code>aspect="length"</code> or "GCfrac", draw vertical lines at the given quantiles of the density estimate. If <code>NULL</code> , no quantile lines will be drawn.
<code>...</code>	Additional argument passed to <code>getColsByBin</code> .

## Value

For `aspect="length"` or "GCfrac", returns a [ggplot](#) object. For `aspect="dinucfreq"`, returns (invisibly) a [Heatmap-class](#) object.

## Examples

```
seqs <- Biostrings::DNASTringSet(
  vapply(1:250, function(i) paste(sample(x = c("A", "C", "G", "T"),
                                         size = round(stats::rnorm(1, 20, 5)),
                                         replace = TRUE), collapse = ""), ""))
bins <- factor(rep(c("a", "b", "c", "d", "e"), each = 50))
plotBinDiagnostics(seqs, bins, aspect = "length")
```

```
plotBinDiagnostics(seqs, bins, aspect = "GCfrac", draw_quantiles = NULL)
plotBinDiagnostics(seqs, bins, aspect = "dinucfreq")
```

---

plotBinHist	<i>Histogram of binned elements.</i>
-------------	--------------------------------------

---

## Description

Plot a histogram of binned elements with binning information.

## Usage

```
plotBinHist(
  x,
  b,
  breaks = 6 * nlevels(b),
  xlab = deparse(substitute(x, env = as.environment(-1))),
  ylab = "Frequency",
  main = "",
  legendPosition = "right",
  legend = NULL,
  legend.cex = NULL,
  ...
)
```

## Arguments

<code>x</code>	A numerical vector with the values used for binning.
<code>b</code>	A factor that groups elements of <code>x</code> into bins (typically the output of <a href="#">bin</a> ).
<code>breaks</code>	A numeric scalar controlling the histogram breaks (passed to <code>geom_hist(..., bins = breaks)</code> ).
<code>xlab, ylab, main</code>	character scalars that set the x-axis label, y-axis label and the main title. Use <code>""</code> to suppress the label.
<code>legendPosition</code>	A character scalar. If not <code>"none"</code> , draw a legend with binning information. The value is used to control the legend position and will be passed to <code>theme(legend.position = legendPosition)</code> .
<code>legend</code>	Deprecated (ignored). Please use <code>legendPosition</code> to control the drawing and position of the legend.
<code>legend.cex</code>	Deprecated (ignored). You can use <a href="#">theme</a> to set legend and other graphical parameters.
<code>...</code>	Further arguments passed to <a href="#">getColsByBin</a> .

## Value

The generated histogram as a `ggplot` object.

## See Also

[getColsByBin](#), [geom\\_histogram](#)

**Examples**

```
set.seed(1)
x <- rnorm(100)
b <- bin(x, "equalN", nElements = 10)
plotBinHist(x, b)
```

plotBinScatter

*Scatter plot (xy-plot) of binned elements.***Description**

Plot a scatter (xy-plot) of binned elements with binning information.

**Usage**

```
plotBinScatter(
  x,
  y,
  b,
  cols = NULL,
  xlab = deparse(substitute(x, env = as.environment(-1))),
  ylab = deparse(substitute(y, env = as.environment(-1))),
  main = "",
  legendPosition = "right",
  legend = NULL,
  legend.cex = NULL,
  ...
)
```

**Arguments**

<code>x</code>	A numerical vector with x values.
<code>y</code>	A numerical vector with y values (the values used for binning).
<code>b</code>	A factor that groups elements of x,y into bins (typically the output of <code>bin(y)</code> ).
<code>cols</code>	NULL or a color vector defining the colors of points. If NULL, the colors will be computed based on b using <code>getColsByBin(b)</code> .
<code>xlab</code>	Label for x-axis.
<code>ylab</code>	Label for y-axis.
<code>main</code>	Main title.
<code>legendPosition</code>	A character scalar. If not "none", draw a legend with binning information. The value is used to control the legend position and will be passed to <code>theme(legend.position = legendPosition)</code> .
<code>legend</code>	Deprecated (ignored). Please use <code>legendPosition</code> to control the drawing and position of the legend.
<code>legend.cex</code>	Deprecated (ignored). You can use <code>theme</code> to set legend and other graphical parameters.
<code>...</code>	Further arguments passed to <code>getColsByBin</code> (only used if <code>cols</code> is NULL).

**Value**

The generated scatter plot as a ggplot object.

**See Also**

[bin](#), [getColsByBin](#), [geom\\_point](#)

**Examples**

```
set.seed(1)
x <- rnorm(100)
y <- rnorm(100)
b <- bin(y, "equalN", nElements = 10)
plotBinScatter(x, y, b)
```

---

plotMotifHeatmaps	<i>Heatmap of motif enrichments.</i>
-------------------	--------------------------------------

---

**Description**

Plot motif enrichments (e.g. significance or magnitude) as a heatmap.

**Usage**

```
plotMotifHeatmaps(
  x,
  which.plots = c("negLog10P", "pearsonResid", "negLog10Padj", "log2enr"),
  width = 4,
  col.enr = c("#053061", "#2166AC", "#4393C3", "#92C5DE", "#D1E5F0", "#F7F7F7",
    "#FDDBC7", "#F4A582", "#D6604D", "#B2182B", "#67001F"),
  col.sig = c("#F0F0F0", "#D9D9D9", "#BDBDBD", "#969696", "#737373", "#525252",
    "#252525", "#000000"),
  col.gc = c("#F7FCF5", "#E5F5E0", "#C7E9C0", "#A1D99B", "#74C476", "#41AB5D", "#238B45",
    "#006D2C", "#00441B"),
  maxEnr = NULL,
  maxSig = NULL,
  highlight = NULL,
  cluster = FALSE,
  show_dendrogram = FALSE,
  show_motif_GC = FALSE,
  show_seqlogo = FALSE,
  show_bin_legend = FALSE,
  width.seqlogo = 1.5,
  use_raster = FALSE,
  na_col = "white",
  doPlot = TRUE,
  ...
)
```

**Arguments**

<code>x</code>	A <a href="#">SummarizedExperiment</a> with numerical matrices (motifs-by-bins) in its <code>assays()</code> , typically the return value of <code>calcBinnedMotifEnrR</code> or <code>calcBinnedMotifEnrHomer</code> .
<code>which.plots</code>	Selects which heatmaps to plot (one or several from "negLog10P", "negLog10Padj", "pearsonResid" and "log2enr").
<code>width</code>	The width (in inches) of each individual heatmap, without legend.
<code>col.enr</code>	Colors used for enrichment heatmap ("pearsonResid" and "log2enr").
<code>col.sig</code>	Colors used for significance heatmaps ("negLog10P" and "negLog10Padj").
<code>col.gc</code>	Colors used for motif GC content (for <code>show_motif_GC = TRUE</code> ).
<code>maxEnr</code>	Cap color mapping at enrichment = <code>maxEnr</code> (default: 99.5th percentile).
<code>maxSig</code>	Cap color mapping at $-\log_{10}$ P value or $-\log_{10}$ FDR = <code>maxSig</code> (default: 99.5th percentile).
<code>highlight</code>	A logical vector indicating motifs to be highlighted.
<code>cluster</code>	If TRUE, the order of transcription factors will be determined by hierarchical clustering of the "pearsonResid" component. Alternatively, an <code>hclust</code> -object can be supplied which will determine the motif ordering. No reordering is done for <code>cluster = FALSE</code> .
<code>show_dendrogram</code>	If <code>cluster != FALSE</code> , controls whether to show a row dendrogram for the clustering of motifs. Ignored for <code>cluster = FALSE</code> .
<code>show_motif_GC</code>	If TRUE, show a column with the percent G+C of the motif as part of the heatmap.
<code>show_seqlogo</code>	If TRUE, show a sequence logo next to each motif label. This will likely only make sense for a heatmap with a low number of motifs.
<code>show_bin_legend</code>	If TRUE, show a legend for the bin labels. If FALSE (default), the bin legend will be hidden.
<code>width.seqlogo</code>	The width (in inches) for the longest sequence logo (shorter logos are drawn to scale).
<code>use_raster</code>	TRUE or FALSE (default). Passed to <code>use_raster</code> of <a href="#">Heatmap</a> .
<code>na_col</code>	"white" (default). Passed to <code>na_col</code> of <a href="#">Heatmap</a> .
<code>doPlot</code>	If TRUE (default), plot the generated heatmap(s) using <code>Reduce(ComplexHeatmap::add_heatmap, heatmapList)</code> . If FALSE, just return the list of heatmap(s) ( <code>heatmapList</code> ) in example before), allowing to modify them further before plotting.
<code>...</code>	Further arguments passed to <a href="#">Heatmap</a> when creating the main heatmaps selected by <code>which.plots</code> . For example, the following will set the font size of the motif names: <code>plotMotifHeatmaps(..., row_names_gp = gpar(fontsize = 12))</code>

**Details**

The heatmaps are created using the **ComplexHeatmap** package and plotted side-by-side.

Each heatmap will be `width` inches wide, so the total plot needs a graphics device with a width of at least `length(which.plots) * width` plus the space used for motif names and legend. The height will be auto-adjusted to the graphics device.

**Value**

A list of `ComplexHeatmap::Heatmap` objects.



## References

Gu, Z. Complex heatmaps reveal patterns and correlations in multidimensional genomic data. *Bioinformatics* 2016.

## See Also

[bin](#), [Heatmap](#)

## Examples

```
se <- readRDS(system.file("extdata",
                          "results.binned_motif_enrichment_LMRs.rds",
                          package = "monalisa"))
i <- which(SummarizedExperiment::assay(se, "negLog10Padj")[, 8] > 4)
plotMotifHeatmaps(se[i, ], which.plots = "pearsonResid",
                  width = 2, show_seqlogo = TRUE)
```

---

plotSelectionProb	<i>Plot selection probabilities of predictors</i>
-------------------	---

---

## Description

This function plots the selection probabilities of predictors (for example the selected motifs), optionally multiplied with either +1 or -1 to give a sense of both the strength and the directionality of the associated effects. The directionality is estimated from the sign of the correlation coefficient between each predictor and the response vector.

## Usage

```
plotSelectionProb(
  se,
  directional = TRUE,
  selProbMin = metadata(se)$stabsel.params.cutoff,
  selProbMinPlot = 0.4,
  showSelProbMin = TRUE,
  selColor = "cadetblue",
  notSelColor = "grey",
  selProbCutoffColor = "firebrick",
  method = c("pearson", "kendall", "spearman"),
  ylimext = 0.2
)
```

## Arguments

se	The SummarizedExperiment object with the results from stability selection (typically returned by <a href="#">randLassoStabSel</a> ).
directional	A logical scalar. If TRUE, selection probabilities are plotted with the sign of the marginal correlation between a predictor and the response.
selProbMin	A numerical scalar in [0,1]. Predictors with a selection probability greater than selProbMin are considered selected and colored by the input from selColor. By default, selProbMin is extracted from the parameters stored in se.

<code>selProbMinPlot</code>	A numerical scalar in [0,1] less than <code>selProbMin</code> . Predictors with a selection probability greater than <code>selProbMinPlot</code> but less than <code>selProbMin</code> are shown as bars with color defined in <code>notSelColor</code> . <code>selProbMinPlot</code> is useful in order to include additional predictors in the barplot, that were not selected according to <code>selProbMin</code> but may be close to that cutoff or are simply nice to visualize alongside the selected predictors. Setting <code>selProbMinPlot = 0</code> will include all predictors.
<code>showSelProbMin</code>	A logical scalar. If TRUE, the value of <code>selProbMin</code> is shown by a horizontal line with the color defined by <code>selProbCutoffColor</code> .
<code>selColor</code>	Color for the selected predictors which have a selection probability greater than <code>selProbMin</code> .
<code>notSelColor</code>	Color for the rest of the (unselected) predictors which will be show in the barplot.
<code>selProbCutoffColor</code>	Color for the line depicting the selection probability cutoff.
<code>method</code>	A character scalar with the correlation method to use in the calculation of predictor-response marginal correlations. One of "pearson", "kendall" or "spearman" (see <a href="#">cor</a> ).
<code>ylimext</code>	A numeric scalar defining how much the y axis limits should be expanded beyond the plotted probabilities to allow for space for the bar labels. This value can be increased if the predictor names above the bars are too long and not showing in the plot.

## Details

This function creates a bar plot with ggplot. Each bar corresponds to a predictor (motif) and the colors correspond to whether or not it was selected. The y-axis shows the selection probabilities (`directional=FALSE`) or selection probabilities with the sign of the marginal correlation to the response (`directional=TRUE`).

## Value

a ggplot2 object.

## Examples

```
## create data set
set.seed(321)
Y <- rnorm(n = 500, mean = 2, sd = 1)
X <- matrix(data = NA, nrow = length(Y), ncol = 50)
for (i in seq_len(ncol(X))) {
  X[,i] <- runif(n = 500, min = 0, max = 3)
}
s_cols <- sample(x = seq_len(ncol(X)), size = 10,
  replace = FALSE)
for (s in s_cols) {
  X[,s] <- X[, s] + (Y + rnorm(500, 0, 4)) * ifelse(s %% 2, -1, 1)
}

## reproducible randLassoStabSel() with 1 core
set.seed(123)
ss <- randLassoStabSel(x = X, y = Y)
plotSelectionProb(ss)
```

---

plotStabilityPaths      *Plot Stability Paths*


---

**Description**

Plot the stability paths of each variable (predictor), showing the selection probability as a function of the regularization step.

**Usage**

```
plotStabilityPaths(
  se,
  selProbMin = metadata(se)$stabsel.params.cutoff,
  selColor = "cadetblue",
  notSelColor = "grey",
  selProbCutoffColor = "firebrick",
  linewidth = 0.5,
  alpha = 1,
  ylim = c(0, 1),
  labelPaths = FALSE,
  labels = NULL,
  labelNudgeX = 8,
  labelSize = 3
)
```

**Arguments**

se	The SummarizedExperiment object resulting from stability selection, by running <a href="#">randLassoStabSel</a> .
selProbMin	A numerical scalar in [0,1]. Predictors with a selection probability greater than selProbMin are shown as colored lines. The color is defined by the col argument.
selColor	Color for the selected predictors which have a selection probability greater than selProbMin.
notSelColor	Color for the rest of the (un-selected) predictors.
selProbCutoffColor	Color for the line depicting the selection probability cutoff.
linewidth	Line width.
alpha	Line transparency of the stability paths.
ylim	Limits for y-axis.
labelPaths	If TRUE, the predictor labels will be shown at the end of the stability paths. The predictor labels given in labels will be shown. If unspecified, the labels corresponding to the selected predictors will be added. If predictors have the same y-value in the last regularization step, the labels will be shown in a random order. One needs to use set.seed to reproduce the plot in this case.
labels	If labelPaths = TRUE, the predictors which should be labelled. If NULL, the selected predictors greater than metadata(se)\$stabsel.params.cutoff will be shown.
labelNudgeX	If labelPaths = TRUE, how much to nudge the labels to the right of the x-axis.
labelSize	If labelPaths = TRUE, the size of the labels.

**Value**

a ggplot2 object.

**See Also**

[stabsel](#)

**Examples**

```
## create data set
Y <- rnorm(n = 500, mean = 2, sd = 1)
X <- matrix(data = NA, nrow = length(Y), ncol = 50)
for (i in seq_len(ncol(X))) {
  X[,i] <- runif(n = 500, min = 0, max = 3)
}
s_cols <- sample(x = seq_len(ncol(X)), size = 10,
  replace = FALSE)
for (i in seq_along(s_cols)) {
  X[,s_cols[i]] <- X[,s_cols[i]] + Y
}

## reproducible randLassoStabSel() with 1 core
set.seed(123)
ss <- randLassoStabSel(x = X, y = Y)
plotStabilityPaths(ss)
```

---

```
prepareHomer
```

---

*Prepare input files for HOMER motif enrichment analysis.*

---

**Description**

For each bin, write genomic coordinates for foreground and background regions into files for HOMER motif enrichment analysis.

**Usage**

```
prepareHomer(
  gr,
  b,
  genomedir,
  outdir,
  motifFile,
  homerfile = findHomer(),
  regionsize = "given",
  Ncpu = 2L,
  verbose = FALSE
)
```

## Arguments

<code>gr</code>	A GRanges object (or an object that can be coerced to one) with the genomic regions to analyze.
<code>b</code>	A vector of the same length as <code>gr</code> that groups its elements into bins (typically a factor).
<code>genomedir</code>	Directory containing sequence files in Fasta format (one per chromosome).
<code>outdir</code>	A path specifying the folder into which the output files (two files per unique value of <code>b</code> ) will be written.
<code>motifFile</code>	A file with HOMER formatted PWMs to be used in the enrichment analysis.
<code>homerfile</code>	Path and file name of the <code>findMotifsGenome.pl</code> HOMER script.
<code>regionsize</code>	The peak size to use in HOMER ("given" keeps the coordinate region, an integer value will keep only that many bases in the region center).
<code>Ncpu</code>	Number of parallel threads that HOMER can use.
<code>verbose</code>	A logical scalar. If TRUE, print progress messages.

## Details

For each bin (unique value of `b`) this functions creates two files in `outdir` (`outdir/bin_N_foreground.tab` and `outdir/bin_N_background.tab`, where `N` is the number of the bin and foreground/background correspond to the ranges that are/are not within the current bin). The files are in the HOMER peak file format (see <http://homer.ucsd.edu/homer/ngs/peakMotifs.html> for details).

In addition, a shell script file is created containing the shell commands to run the HOMER motif enrichment analysis.

## Value

The path and name of the script file to run the HOMER motif enrichment analysis.

## Examples

```
# prepare genome directory (here: one dummy chromosome)
genomedir <- tempdir()
dir.create(genomedir)
writeLines(c(">chr1", "ATGCATGCATCGATCGATCGATCGTACGTA"),
           file.path(genomedir, "chr1.fa"))

# prepare motif file, regions and bins
motiffile <- tempdir()
dumpJaspar(filename = motiffile, pkg = "JASPAR2020",
           opts = list(ID = c("MA0006.1")))
gr <- GenomicRanges::GRanges("chr1", IRanges::IRanges(1:4, width = 4))
b <- bin(1:4, nElements = 2)

# create dummy file (should point to local Homer installation)
homerfile <- file.path(tempdir(), "findMotifsGenome.pl")
writeLines("dummy", homerfile)

# run prepareHomer
outdir <- tempdir()
prepareHomer(gr = gr, b = b, genomedir = genomedir,
             outdir = outdir, motifFile = motiffile,
             homerfile = homerfile, verbose = TRUE)
```

```
list.files(outdir)

# clean up example
unlink(c(genomedir, motiffile, homerfile, outdir))
```

---

randLassoStabSel

*Randomized Lasso Stability Selection*


---

## Description

This function runs randomized lasso stability selection as presented by Meinshausen and Bühlmann (2010) and with the improved error bounds introduced by Shah and Samworth (2013). The function uses the [stabsel](#) function from the `stabs` package, but implements the randomized lasso version.

## Usage

```
randLassoStabSel(
  x,
  y,
  weakness = 0.8,
  cutoff = 0.8,
  PFER = 2,
  mc.cores = 1L,
  glmnet.args = list(),
  ...
)
```

## Arguments

<code>x</code>	The predictor matrix.
<code>y</code>	The response vector.
<code>weakness</code>	Value between 0 and 1 (default = 0.8). It affects how strict the method will be in selecting predictors. The closer it is to 0, the more stringent the selection. A weakness value of 1 is identical to performing lasso stability selection (not the randomized version).
<code>cutoff</code>	Value between 0 and 1 (default = 0.8) which is the cutoff for the selection probability. Any variable with a selection probability that is higher than the set cutoff will be selected.
<code>PFER</code>	Integer (default = 2) representing the absolute number of false positives that we allow for in the final list of selected variables. For details see Meinshausen and Bühlmann (2010).
<code>mc.cores</code>	Integer (default = 1) specifying the number of cores to use in <a href="#">mclapply</a> , which is the default way <a href="#">stabsel</a> does parallelization.
<code>glmnet.args</code>	Named list with additional arguments to the internal <code>.glmnetRandomizedLasso</code> function (beyond <code>x</code> , <code>y</code> and <code>weakness</code> , which are determined automatically, and <code>q</code> , which should not be specified (it will be determined from <code>cutoff</code> and <code>PFER</code> ). The available arguments to <code>.glmnetRandomizedLasso</code> are the same as the ones for <a href="#">glmnet.lasso</a> . A typical use case would be to define the <code>family</code> argument to <a href="#">glmnet</a> .
<code>...</code>	Additional parameters that can be passed on to <a href="#">stabsel</a> .

## Details

Randomized lasso stability selection runs a randomized lasso regression several times on subsamples of the response variable and predictor matrix.  $N/2$  elements from the response variable are randomly chosen in each regression, where  $N$  is the length of the vector. The corresponding section of the predictor matrix is also chosen, and the internal `.glmnetRandomizedLasso` function is applied. Stability selection results in selection probabilities for each predictor. The probability of a specific predictor is the number of times it was selected divided by the total number of subsamples that were done (total number of times the regression was performed).

We made use of the `stabs` package that implements lasso stability selection, and adapted it to run randomized lasso stability selection.

## Value

A `SummarizedExperiment` object where the rows are the observations and the columns the predictors (same dimnames as the predictor matrix `x`). It contains:

**assays** :

**x** : the predictor matrix.

**rowData** : a `DataFrame` with columns:

**y** : the response vector.

**colData** : a `DataFrame` with columns:

**selProb** : the final selection probabilities for the predictors (from the last regularization step).

**selected** : logical indicating the predictors that made the selection with the specified cutoff.

**selAUC** : the normalized area under the selection curve (mean of selection probabilities over regularization steps).

**reg'i'** : columns containing the selection probabilities for regularization step  $i$ .

**metadata** : a list of output returned from `stabsel` and `randLassoStabSel`:

**stabsel.params.cutoff** : probability cutoff set for selection of predictors (see `stabsel`).

**stabsel.params.selected** : elements with maximal selection probability greater cutoff (see `stabsel`).

**stabsel.params.max** : maximum of selection probabilities (see `stabsel`).

**stabsel.params.q** : average number of selected variables used (see `stabsel`).

**stabsel.params.PFER** : (realized) upper bound for the per-family error rate (see `stabsel`).

**stabsel.params.specifiedPFER** : specified upper bound for the per-family error rate (see `stabsel`).

**stabsel.params.p** : the number of effects subject to selection (see `stabsel`).

**stabsel.params.B** : the number of subsamples (see `stabsel`).

**stabsel.params.sampling.type** : the sampling type used for stability selection (see `stabsel`).

**stabsel.params.assumption** : the assumptions made on the selection probabilities (see `stabsel`).

**stabsel.params.call** : `stabsel` the call.

**randStabsel.params.weakness** : the weakness parameter in the randomized lasso stability selection.

## References

N. Meinshausen and P. Bühlmann (2010), Stability Selection, *Journal of the Royal Statistical Society: Series B (Statistical Methodology)*, **72**, 417–73.

R.D. Shah and R.J. Samworth (2013), Variable Selection with Error Control: Another Look at Stability Selection, *Journal of the Royal Statistical Society: Series B (Statistical Methodology)*, **75**, 55–80.

B. Hofner, L. Boccutto, and M. Göker (2015), Controlling False Discoveries in High-Dimensional Situations: Boosting with Stability Selection, *BMC Bioinformatics*, **16** 144.

## See Also

[stabsel](#)

## Examples

```
## create data set
Y <- rnorm(n = 500, mean = 2, sd = 1)
X <- matrix(data = NA, nrow = length(Y), ncol = 50)
for (i in seq_len(ncol(X))) {
  X[,i] <- runif(n = 500, min = 0, max = 3)
}
s_cols <- sample(x = seq_len(ncol(X)), size = 10,
  replace = FALSE)
for (i in seq_along(s_cols)) {
  X[,s_cols[i]] <- X[,s_cols[i]] + Y
}

## reproducible randLassoStabSel() with 1 core
set.seed(123)
ss <- randLassoStabSel(x = X, y = Y)

## reproducible randLassoStabSel() in parallel mode
## (only works on non-windows machines)
if (.Platform$OS.type == "unix") {
  RNGkind("L'Ecuyer-CMRG")
  set.seed(123)
  ss <- randLassoStabSel(x = X, y = Y, mc.preschedule = TRUE,
    mc.set.seed = TRUE, mc.cores = 2L)
}
```

---

sampleRandomRegions	<i>Sample random regions of fixed length.</i>
---------------------	---

---

## Description

Sample random regions from the mappable parts of the genome with a given fraction from CpG islands.

## Usage

```
sampleRandomRegions(allowedRegions = NULL, N = 100L, regWidth = 200L)
```



**Arguments**

allowedRegions	An unstranded GRanges object of the "allowed" of the genome, usually the mappable regions.
N	Number of regions to sample.
regWidth	Region width.

**Details**

In order to make the results deterministic, set the random number seed before calling `sampleRandomRegions` using `set.seed`.

**Value**

A GRanges object with randomly sampled mappable regions of width `regWidth` with `fractionCGI` coming from CpG islands.

**Examples**

```
regs <- GenomicRanges::GRanges(
  seqnames = rep(c("chr1", "chr2"), each = 2),
  ranges = IRanges::IRanges(start = 1:4, end = 5:8))
set.seed(123)
sampleRandomRegions(regs, N = 2, regWidth = 3L)
```

---

seqLogoGrob	<i>Create a simple sequence logo grob.</i>
-------------	--

---

**Description**

Create a simple sequence logo grob (grid-graphics object) for a transcription factor from a position frequency matrix. The logo drawing code is a simplified version from [seqLogo](#) and for example can be used to embedd sequence logos within other plots.

**Usage**

```
seqLogoGrob(x, xmax = NULL, ymax = 2, xjust = c("left", "center", "right"))
```

**Arguments**

x	A <a href="#">PFMatrix</a> object
xmax	A numeric scalar with the maximal width for the logo (in base-pairs). A value of NULL will scale the logo to the full width of the viewport.
ymax	A numeric scalar with the maximal height for the logo (in bits) A value of NULL will scale the logo to the full height of the viewport.
xjust	A character scalar specifying the horizontal adjustment of the sequence log within the viewport; one of "left", "center" or "right".

**Value**

A polygon grob.

**See Also**

[seqLogo](#) for the original, more flexible version of this function.

**Examples**

```
if (require(JASPAR2020) && require(TFBSTools) && require(gridExtra)) {  
  pfm1 <- getMatrixByID(JASPAR2020, "MA0139")  
  pfm2 <- getMatrixByID(JASPAR2020, "MA0531")  
  
  g1 <- seqLogoGrob(pfm1)  
  g2 <- seqLogoGrob(pfm2)  
  
  gridExtra::grid.arrange(g1, g2)  
}
```

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